

Cannabinoids, the endocannabinoid system, and pain: a review of preclinical studies

David P. Finn^{a,*}, Simon Haroutounian^b, Andrea G. Hohmann^c, Elliot Krane^d, Nadia Soliman^e, Andrew S.C. Rice^e

Abstract

This narrative review represents an output from the International Association for the Study of Pain's global task force on the use of cannabis, cannabinoids, and cannabis-based medicines for pain management, informed by our companion systematic review and meta-analysis of preclinical studies in this area. Our aims in this review are (1) to describe the value of studying cannabinoids and endogenous cannabinoid (endocannabinoid) system modulators in preclinical/animal models of pain; (2) to discuss both pain-related efficacy and additional pain-relevant effects (adverse and beneficial) of cannabinoids and endocannabinoid system modulators as they pertain to animal models of pathological or injury-related persistent pain; and (3) to identify important directions for future research. In service of these goals, this review (1) provides an overview of the endocannabinoid system and the pharmacology of cannabinoids and endocannabinoid system modulators, with specific relevance to animal models of pathological or injury-related persistent pain; (2) describes pharmacokinetics of cannabinoids in rodents and humans; and (3) highlights differences and discrepancies between preclinical and clinical studies in this area. Preclinical (rodent) models have advanced our understanding of the underlying sites and mechanisms of action of cannabinoids and the endocannabinoid system in suppressing nociceptive signaling and behaviors. We conclude that substantial evidence from animal models supports the contention that cannabinoids and endocannabinoid system modulators hold considerable promise for analgesic drug development, although the challenge of translating this knowledge into clinically useful medicines is not to be underestimated.

Keywords: Cannabinoid₁ (CB₁) receptor, Cannabinoid₂ (CB₂) receptor, Endocannabinoid, Chronic pain, Neuropathic pain, Inflammatory pain, Nociception, Rats, Mice, Behavior

1. Introduction

Cannabis, cannabis extracts or oils, individual cannabinoid substances, and modulators of the endogenous cannabinoid (endocannabinoid) system have all been suggested as therapeutic agents for pain management.^{75,94,193} The primary drivers for interest in these agents include (1) major scientific advances in our understanding of the biology of the endocannabinoid system and pharmacology of cannabinoids; (2) development and regulatory approval of cannabis-based (or cannabis-derived) medicines (eg, Nabiximols/Sativex and Epidiolex); and (3) regulatory changes that permit use of cannabis for medical purposes, including pain management, following advocacy by patients and public support.

However, the latter development has proven controversial amongst scientists, patients, and healthcare professionals, with uncertainty over whether the current evidence base for efficacy and safety justifies change in legislation of cannabis for medical use, including pain management. In 2018, the International Association for Study of Pain (IASP) established a Presidential Task Force on Cannabis and Cannabinoids Analgesia. The present narrative review represents an output from work package 1 of the IASP task force, which was focused on basic science, including definition of terminology, overview of the endogenous cannabinoid (endocannabinoid) system, compound classification, pharmacology, and assessment of pain-related efficacy and additional pain-relevant effects (adverse and beneficial) in preclinical laboratory animal studies.

The aims of this review are:

- (1) To provide commentary on the efficacy and side effects of cannabinoids and endocannabinoid system modulators as they pertain to animal models of pathological pain (the term "animal model of pain" is not a universally agreed descriptor, but given that it is common usage, we will use it herein as shorthand. It is also worth noting that there is a distinction between "model," which reflects the underlying disease or injury, and the pain-associated outcome measures used in evaluating such models), informed by our companion systematic review and meta-analysis of preclinical studies in this area.²¹⁵
- (2) To discuss the value of studying cannabinoids and endocannabinoid system modulators in preclinical/animal models of pain, as well as discrepancies between preclinical and clinical studies in this area, and

Sponsorships or competing interests that may be relevant to content are disclosed at the end of this article.

^a Pharmacology and Therapeutics, School of Medicine, Galway Neuroscience Centre and Centre for Pain Research, Human Biology Building, National University of Ireland Galway, Galway, Ireland, ^b Department of Anesthesiology and Washington University Pain Center, Washington University in St. Louis School of Medicine, St. Louis, MO, United States, ^c Psychological and Brain Sciences, Program in Neuroscience, and Gill Center for Biomolecular Science, Indiana University, Bloomington, IN, United States, ^d Departments of Anesthesiology, Perioperative, and Pain Medicine, & Pediatrics, Stanford University School of Medicine, Stanford, CA, United States, ^e Pain Research, Department of Surgery & Cancer, Faculty of Medicine, Imperial College London, United Kingdom

*Corresponding author. Address: Pharmacology and Therapeutics, School of Medicine, Human Biology Building, National University of Ireland Galway, University Rd, Galway, H91 W5P7, Ireland. Tel.: +353 (0)91 495280. E-mail address: david.finn@nuigalway.ie (D.P. Finn).

PAIN 162 (2021) S5–S25

© 2021 International Association for the Study of Pain

<http://dx.doi.org/10.1097/j.pain.0000000000002268>

(3) Provide suggestions for future research directions.

In service of the above objectives, we:

- (1) Clearly define terminology used in this field.
- (2) Provide an overview of the endocannabinoid system.
- (3) Review the classification, chemistry, and pharmacology of cannabinoids, cannabis-based medicines (CBM), and endocannabinoid system modulators, including structure–activity relationships and pharmacokinetics in rodents and humans.

2. Terminology and definitions

One factor that has hampered public debate (and sometimes scientific/medical debate) on the topic of cannabis and cannabinoids for pain management is the inappropriate, inconsistent, or unclear use of terminology. For example, the terms “cannabis,” “cannabinoids,” and “cannabis-based medicines (CBM)” are often used interchangeably or conflated, both within public discourse and the media, and within the scientific literature (eg, pooling of data relating to cannabis and individual cannabinoids in meta-analyses and systematic reviews of clinical efficacy). Cannabis refers to the whole plant, or to its parts, and must be clearly distinguished from cannabinoid ligands (cannabinoids) that are either plant-derived natural (phytocannabinoids), synthetic, or semisynthetic, but always chemically defined, single-entity compounds usually having affinity for and activity at cannabinoid receptors. Thus, cannabis, single-entity cannabinoid compounds, and modulators of the endocannabinoid system should not be used synonymously or conflated within the same preclinical or clinical investigation for either efficacy or side effects. Furthermore, when considering or debating the merits, or otherwise, of cannabis, due regard must be given to the fact that cannabis is highly heterogeneous with many different chemical constituents and that a multitude of different strains of the plant exist, all containing different amounts of phytocannabinoids,^{12,103} most particularly Δ^9 -tetrahydrocannabinol (THC) and cannabidiol (CBD). Tetrahydrocannabinol, first isolated in the 1960s, is the primary psychoactive constituent of *Cannabis sativa*.^{83,162} Tetrahydrocannabinol has psychoactive properties, and its pharmacological effects are attributable to agonist activity at both cannabinoid type 1 (CB₁) and type 2 (CB₂) receptors.^{44,61,148,234} Cannabidiol, by contrast, does not have appreciable agonist activity at CB₁ receptors (but may be a negative allosteric modulator of CB₁) and lacks the psychoactivity profile of THC.^{22,68,222} In addition to phytocannabinoids, the cannabis plant also contains a large number of terpenes, flavonoids, and other compounds, which, themselves, may be pharmacologically active but remain understudied.

Cannabis-based medicines are registered medicinal cannabis extracts, approved by regulatory authorities, with well-defined, standardized THC/CBD content, and with no, or only trace levels of, terpenes, flavonoids, and other compounds. Examples of CBMs are nabiximols (Sativex), approved in some countries for adjunctive treatment of spasticity (and in some jurisdictions neuropathic pain) in multiple sclerosis, and CBD extract (Epidiolex), indicated for treatment of childhood epilepsy, which has only minor or trace levels of other phytocannabinoids. These medicinal products with regulatory approval should be distinguished from the so-called cannabis or CBD oils or extracts, which are numerous, typically sold in health food stores, pharmacies, cannabis dispensaries, or over the internet, but often have uncertain and/or unverified THC/CBD content. Synthetic THC (eg, dronabinol) and the synthetic THC analogue nabilone are also approved for indications such as anorexia and weight loss in AIDS, and chemotherapy-induced nausea and

vomiting, and available on special prescription in many countries.

Table 1 provides a glossary of terminology and definitions.

3. The endocannabinoid system

The discovery of THC in the 1960s as the psychoactive constituent of *Cannabis* inspired research into its pharmacology and mechanism of action. However, it was not until the late 1980s/early 1990s that the cannabinoid receptors were discovered (**Fig. 1**). The first cannabinoid receptor to be discovered, cloned, and characterised was the CB₁ receptor, initially in rat brain^{57,159} and subsequently localized in human brain.⁶⁸ In 1993, a second cannabinoid receptor, CB₂, was cloned and characterized in a human promyelocytic leukaemic cell line and in rat spleen.¹⁶⁷ Both CB₁ and CB₂ belong to the superfamily of seven-transmembrane domain, G-protein-coupled receptors (G_{i/o} coupled). The existence of cannabinoid receptors, highly conserved across species, implied the existence of endogenous cannabis-like molecules (endocannabinoids) that bind to and modulate cannabinoid receptors.

Research efforts sought to identify the endogenous ligands that bind to and modulate mammalian cannabinoid receptors. Two endocannabinoids, *N*-arachidonylethanolamide (anandamide [AEA])⁵⁸ and 2-arachidonoyl glycerol (2-AG),^{161,220} were discovered. The endocannabinoid system is an important physiological system, composed of CB₁ and CB₂ receptors, their endogenous ligands, AEA and 2-AG, and the enzymes responsible for the synthesis and degradation of the endocannabinoids. Anandamide and 2-AG are the best characterised endocannabinoids. However, there are several other endogenous ligands with affinity and activity at cannabinoid receptors, including 2-AG ether (noladin ether), virodhamine, *N*-arachidonoyl dopamine, and others.^{10,59,63,191,192}

The CB₁ receptor is the most highly expressed G-protein-coupled receptor subtype in the central nervous system (CNS).^{88,104,191} Within the brain, CB₁ is found in high density in the basal ganglia, as well as in key components of the descending pain pathway and the stress/fear/anxiety circuitry. The CB₁ receptor is localized to most other tissues and organs of the body. Of relevance to pain, in addition to their supraspinal localisation, CB₁ receptors are expressed in the dorsal horn of the spinal cord, synthesized in dorsal root ganglion cells, and transported in peripheral nerves.^{21,74,108–111,178} Immunohistochemical studies have localized CB₁ to dorsal root ganglia and identified CB₁ receptors on primary afferent neurons.^{21,110,111} Cannabinoid type 2 receptors are mainly expressed in the periphery, with particularly high density on cells and tissues of the immune system.^{11,167} Although localisation of CB₂ to otherwise naive CNS remains controversial, CB₂ can be induced in the CNS in response to injury or pathophysiological states (for review, see Refs. 99, 134, 205, and 240). Being G_{i/o}-protein-coupled receptors, CB₁ and CB₂ are negatively coupled to adenylyl cyclase,^{116–118} and positively coupled to mitogen-activated protein kinase.¹⁸ Within neurons, CB₁ activation results in inhibition of N- and P/Q-type voltage-activated Ca²⁺ channels, and induction of inwardly rectifying K⁺ currents, with consequent inhibition of neurotransmitter release.⁵⁵ Cannabinoids, including endocannabinoids, phytocannabinoids, and synthetic cannabinoids, may potentially also act at other non-CB₁/non-CB₂ receptors, including the transient receptor potential cation channel subfamily V member 1 (TRPV1; also known as the capsaicin or vanilloid receptor VR1), peroxisome proliferator-activated receptors, and G-protein-coupled receptors such as *GPR55* and *GPR119*.^{6,23,179}

Table 1
Terminology and Definitions (Adapted from Soliman et al., 2019, after modification from Hauser et al., 2018).

Term	Definition	Examples/typical products
(Herbal) cannabis	The whole plant or parts or material from the plant (eg, flowers, buds, resin, leaves)	<i>Cannabis sativa</i> , hashish
Medical or medicinal cannabis	The term “medical/medicinal cannabis” (or “medical/medicinal marijuana”) is used for cannabis plants, plant material, or full plant extracts used for medical purposes.	Bedrocan, Bedrobinol, tilray 10THC/10CBD
Cannabis-based (or cannabis-derived) medicines	Medicinal cannabis extracts or products with regulatory approval for marketing as a therapeutic with defined and standardized THC and/or CBD content.	Nabiximols (Sativex), dronabinol (Marinol), Epidiolex
Cannabinoids	Cannabinoids are biologically active constituents of cannabis, or synthetic compounds, usually having affinity for and activity at cannabinoid receptors.	THC, CBD, CP55,940, WIN55,212-2, HU210, nabilone
Phytocannabinoid	A cannabinoid found in cannabis plants or purified/extracted from plant material	THC, CBD
Endocannabinoid	An endogenous ligand found in the body of humans and other animals and which has affinity for, and activity at, cannabinoid receptors	Anandamide, 2-AG
Modulators that decrease endocannabinoid system activity	Directly block cannabinoid receptors or reduce signalling indirectly via impeding action of endogenous ligand through actions at a distinct site	Cannabinoid receptor antagonists (rimonabant [SR141716A], AM251, SR144528, AM630), negative allosteric modulators (PSNCBAM-1), DAGL inhibitors (RHC80267)
Modulators that increase or enhance endocannabinoid system activity	In addition to individual phytocannabinoids, cannabis-derived or cannabis-based medicines, and cannabis extracts, other pharmacological approaches under development for manipulation of the endocannabinoid system include selective synthetic cannabinoid receptor agonists, inhibitors of the catabolism (eg, fatty acid amide hydrolase [FAAH] inhibitors), transport (eg, fatty acid-binding protein [FABP] inhibitors) or reuptake of endocannabinoids, or positive allosteric modulators of cannabinoid receptor signalling.	FAAH inhibitors (PF-04457845, URB597, URB937), anandamide transport inhibitors (AM404, VDM11), MGL inhibitors (URB602, JZL184, MJN110), positive allosteric modulators of the CB ₁ receptor (ZCZ011, GAT211)

CBD, cannabidiol; CB₁, cannabinoid type 1; DAGL, diacylglycerol lipase; FABP, fatty acid-binding protein; THC, Δ⁹-tetrahydrocannabinol; 2-AG, 2-arachidonoyl glycerol; MGL, monoacylglycerol lipase.

Mechanisms underlying endocannabinoid biosynthesis, signaling, and degradation are quite well understood. Biosynthesis of AEA involves its formation from the precursor N-

arachidonoylphosphatidylethanolamine (NAPE), catalysed by the hydrolytic activity of the phospholipase D enzyme known as N-acylphosphatidylethanolamine-hydrolyzing phospholipase D

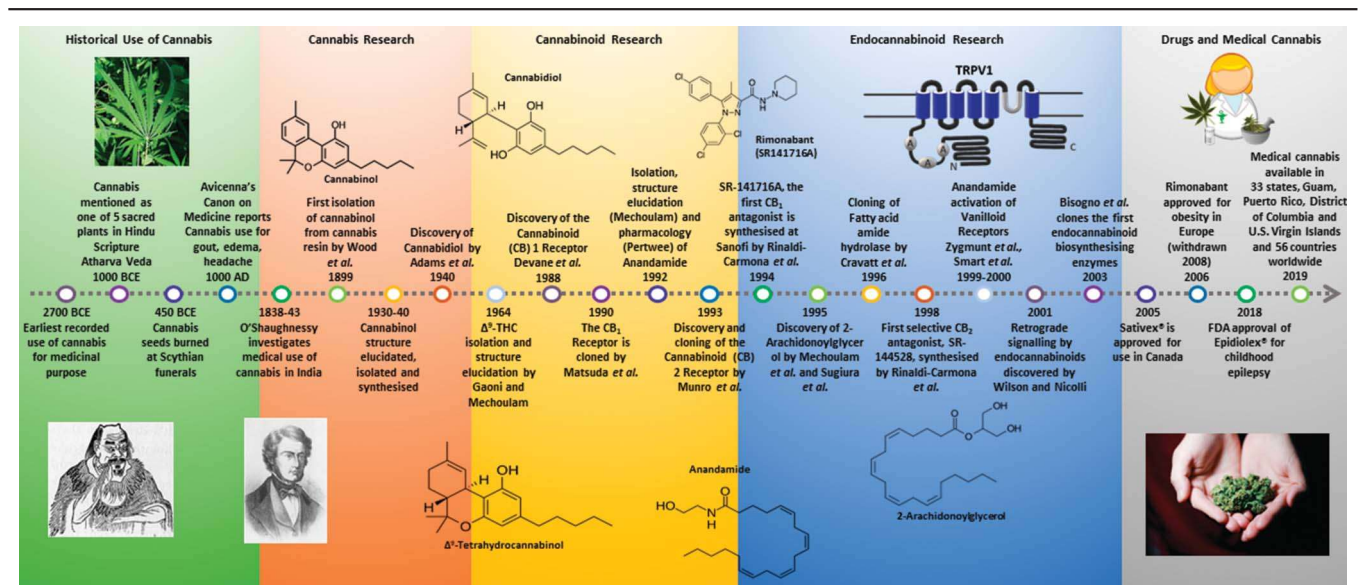


Figure 1. A historical timeline of key milestones in cannabis and cannabinoid research.

(for review, see Refs. 13, 30, 31, and 62). 2-arachidonoyl glycerol is synthesized by conversion of 1,2-diacylglycerol to 2-AG by diacylglycerol lipases (for review, see Refs. 59, 135, 219, and 227). Anandamide is primarily degraded to arachidonic acid and ethanolamine by the enzyme fatty acid amide hydrolase (FAAH), located in the endoplasmic reticulum of postsynaptic neuron^{46,59} (for review, see Refs. 86 and 182). Fatty acid amide hydrolase also catabolizes other *N*-acylethanolamines including *N*-palmitoylethanolamide (PEA) and *N*-oleoylethanolamide, which themselves do not have appreciable activity at CB₁ or CB₂ receptors but may elevate levels of AEA through substrate competition at FAAH.¹³³ 2-arachidonoyl glycerol is primarily metabolized to arachidonic acid and glycerol by the presynaptic enzyme monoacylglycerol lipase (MGL),^{65,227} with other enzymes including FAAH, ABHD6, and ABHD12 accounting for a modest (ie, <10%) degree of 2-AG catabolism.^{15,89} Within the nervous system, newly synthesized endocannabinoids leave the postsynaptic neuron to exert their effects on CB₁ receptors expressed on presynaptic nerve terminals in a signaling process known as retrograde neurotransmission. Further work is required to better understand the mechanisms by which endocannabinoids are transported within cells and across cell membranes.

As a lipid-signalling system whose components are expressed widely across the body, the endocannabinoid system plays a key role in the regulation of a wide array of physiological processes including metabolism, mood, motor function, appetite, cardiovascular control, stress response, gastrointestinal tract function, developmental biology, cell fate, immune and inflammatory response, endocrine function, neurotransmission, and pain (for review, see Refs. 60, 183, and 184). In the context of nociception and pain, key components of the endocannabinoid system are expressed throughout nociceptive pathways (**Fig. 2**): in the periphery on primary afferent neurons, in the dorsal horn of the spinal cord, and in multiple supraspinal regions of the brain associated with pain perception and modulation (for review, see Refs. 96, 99, 196, 207, 217, and 240). As a result, targeting the endocannabinoid system via enhancement of the levels of endogenous cannabinoids (eg, with FAAH or MGL inhibitors) or exogenous cannabinoid ligands (eg, CB₁ or CB₂ receptor agonists) can reduce nociceptive transmission at all 3 of these neuroanatomical levels. Glial cells, which express components of the endocannabinoid system, represent another substrate through which cannabinoids or endocannabinoid system modulators may regulate pain through neuroimmune interactions^{29,157} (for review, see Refs. 233, 238, and 242). Preclinical research indicates that endocannabinoids are synthesized on-demand in postsynaptic neurons in response to stress or pain and produce short-term antinociceptive effects via presynaptic inhibitory CB₁ receptors.^{92,113,144,181,218,239} Endocannabinoids are implicated in control of pain initiation^{32,38} and play an important role in the resolution of tonic pain and in stress-induced and fear-conditioned analgesia in rodents.^{27,28,77,92,113} In animal models of pathological pain, the endocannabinoid system exhibits adaptive changes or plasticity (eg, altered cannabinoid receptor expression/functionality and endocannabinoid levels) depending on the model and anatomical site under investigation.^{100,200,206,240,241} These findings support the contention that the endocannabinoid system may represent a viable therapeutic target for chronic pain. This view is supported by numerous pharmacological studies demonstrating efficacy of cannabinoids or modulators of the endocannabinoid system in animal models of pathological or injury-related pain. These latter studies have been reviewed and analysed in our systematic review and meta-analysis of the preclinical literature published in

this special issue of PAIN.²¹⁵ Herein, we include a summary of the key findings of this systematic review and extended commentary on some elements not discussed in detail within the systematic review itself.

4. Classification of cannabinoids and endocannabinoid system modulators of relevance to pain

Common pharmacological tools used to manipulate the endocannabinoid system are summarized in **Table 2**.

5. Antinociceptive efficacy of cannabinoids in animal models of pathological or injury-related persistent pain

The antinociceptive efficacy of cannabinoids and endocannabinoid system modulators has been reviewed extensively.^{99,196,206,217,240,241} Thus, herein, we instead provide a concise commentary on the efficacy of cannabinoids and endocannabinoid system modulators in animal models of pathological or injury-related persistent pain that is uniquely informed by our companion systematic review and meta-analysis of preclinical studies in this area.²¹⁵ We summarise the key findings and elaborate further on some of their implications for the field.

Our companion systematic review of laboratory animal studies that used models/conditions associated with persistent pain and reported a pain-relevant outcome measure identified 473 published reports of which 374 reported data that could be included in a meta-analysis.²¹⁵ Data from 6479 rats and 6876 mice, respectively, were included, reflecting 864 and 677 experimental comparisons, respectively. This is a very large data set by preclinical standards. No studies in other species were found. Ninety-nine studies (~20%) were excluded from the meta-analysis because the methods and/or results were not reported in sufficient detail to permit extractable data to be included in a meta-analysis.

Overall, the data support the hypothesis of cannabinoid-mediated analgesia. The overall effect size (Hedges' *G* standardised mean difference) was 1.32 [*Q* = 4101.26, *df* 1543, *P* < 0.0001, *I*² = 61.58%] (**Fig. 3**). The models used reflect a range of conventional and diverse inflammatory and neuropathy paradigms, particularly surgically induced nerve injury for the latter. With regard to outcome measures, in common with other preclinical behavioral pain studies, limb withdrawal measures evoked by sensory stimuli were by far the most frequently reported outcome measure, with very few reports of complex behavioral assessments. Lack of consensus on predictive validity of such measures limits the degree to which the presence of pain, and thus pain relief from the intervention, can be inferred.^{190,202} This contrasts with the clinical trial literature where patient-reported pain intensity is the predominant metric. The formalin test also features strongly in the data; this model of tonic inflammatory pain entails measuring several spontaneous or nonevoked nociceptive behaviors, for example, licking, lifting, flinching, rearing, and guarding, to generate a "combined or composite pain score," which thereby affords a greater degree of confidence regarding the impact of pain on the animal's behavior.^{1,69,224}

Small-molecule CB₁ and CB₂ receptor agonists and non-selective cannabinoid receptor agonists (including THC) were the most frequently assessed interventions. Fatty acid amide hydrolase inhibitors and peroxisome proliferator-activated

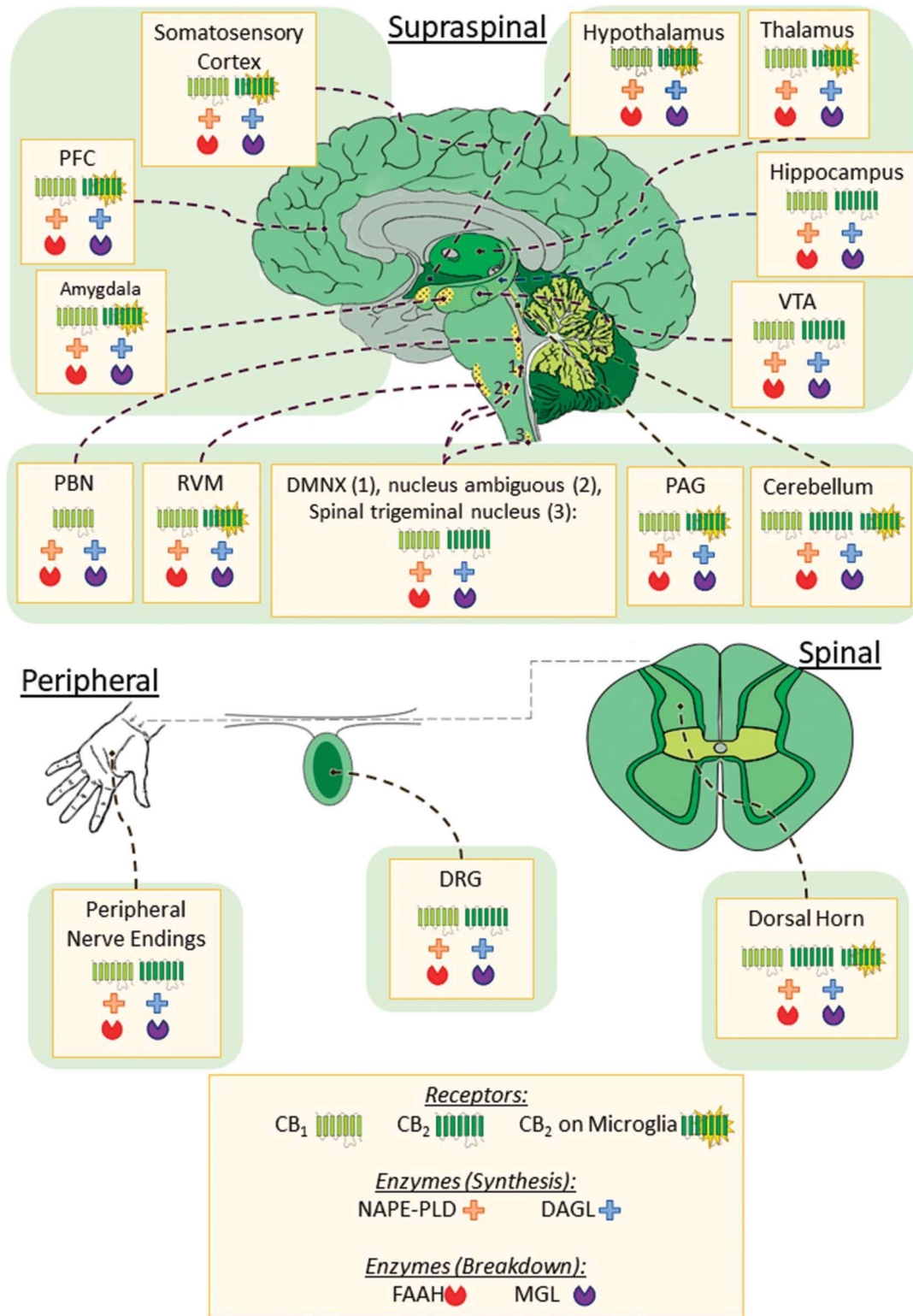


Figure 2. Distribution of the cannabinoid receptors and enzymes associated with endocannabinoid synthesis and degradation in pain pathways. The endocannabinoid system is widely distributed throughout regions associated with pain processing and modulation in the brain, spinal cord, and periphery, most particularly the CB₁ receptor and the enzymes responsible for endocannabinoid synthesis (NAPE-PLD and DAGL) and degradation (FAAH and MGL). CB₂ receptors are less abundant in the brain and are primarily located on microglia; however, studies have shown expression of CB₂ on neurons in the VTA, and several discrete nuclei of the brainstem. CB₁ is also expressed on microglia at a lower level than CB₂, and effects of CB₁ on microglia may be mediated by neuronal CB₁. Therefore, this was not depicted above. In the DRG and periphery, both CB₁ and CB₂ receptors can be found on neurons (and glia in the dorsal horn), as well as the endocannabinoid enzymes. PFC, prefrontal cortex; VTA, ventral tegmental area; PAG, periaqueductal gray; RVM, rostral ventromedial medulla; PBN, parabrachial nucleus; DMNX, dorsal motor nucleus of the vagus nerve; DRG, dorsal root ganglion; CB, cannabinoid receptor; NAPE-PLD, N-acylphosphatidylethanolamine-hydrolyzing phospholipase D; DAGL, diacylglycerol lipase; FAAH, fatty acid amide hydrolase; MGL, monoacylglycerol lipase.

Downloaded from http://journals.lww.com/pain by BMDMFepHkav1ZEoum1tIQIN4a+kLLHEZgbsH04XIM0hCwWCX1AW nYQpjlIcHHD33D00dRv17TSfH4C3VC1y0abggQZXdggZ2MwZLq= on 03/29/2024

Table 2
Cannabinoid ligands/preparations and pharmacological tools.

Natural cannabinoid ligands/ Synthetic analogues	CB ₁ -selective agonists
*Δ ⁹ -THC (dronabinol)	ACEA
*CBD	Met-F-AEA AZ11713908 (peripherally restricted with CB ₂ inverse agonist properties)
	Mixed CB ₁ /CB ₂ agonists
*Cannabis	BAY59-3074
*eCBD	CP55,940
*Nabilone (Δ ⁹ -THC Analogue)	*CT-3 (ajulemic acid)
*Nabiximols (oral-mucosal spray, Δ ⁹ -THC:CBD, 2.7:2.5 mg)	HU-210
Endocannabinoids	WIN55,212-2
AEA	CB ₂ -selective agonists
2-AG	AM1241
Endocannabinoid modulators	AM1714
CB ₁ -Positive Allosteric Modulators	AM1710
GAT211	A-796260
GAT229	A-836339
ZCZ011	*GW842166X
Uptake Inhibitors	*HU308
AM404	JWH015
LY2318912	JWH133
VDM11	LY2828360
OMDM132	MDA7
UCM-707	MDA19
FAAH inhibitors	CB ₁ antagonists
OL135	AM251
*PF-00457845	AM281
URB597	AM6545 (peripherally restricted)
URB937 (peripherally restricted)	SR141716
MGL inhibitors	CB ₂ antagonists
JZL184	AM630
URB602 (local only)	SR144528
MJN110	Fatty acids that do not bind CBRs
Dual FAAH-MGL inhibitors	NaGly
JZL195	PEA
SA57	L-29
FABP5 inhibitors	
SBF126	

* Compounds used or tested clinically; Adapted and updated from Hohmann and Rice (2013) Textbook of Pain sixth Edition and Rahn and Hohmann (2009) Neurotherapeutics 6:713-3.

AEA, anandamide; CBD, cannabidiol; CB₁, cannabinoid type 1; CB₂, cannabinoid type 2; eCBD, high CBD cannabis; Δ⁹-THC, Δ⁹-tetrahydrocannabinol; FAAH, fatty acid amide hydrolase; FABP, fatty acid-binding protein; FLAT, FAAH-like anandamide transporter; MGL, monoacylglycerol lipase; N-arachidonoyl glycine, NaGly; PEA, palmitoylethanolamide 2-AG, 2-arachidonoyl glycerol.

receptor-α agonists (in particular, palmitoylethanolamide; PEA) were also frequently evaluated. In general, studies demonstrated antinociceptive efficacy, as measured predominantly by attenuation of injury-/inflammation-associated hypersensitivity in evoked limb withdrawal (Fig. 4).

The differences between the effect sizes of the different interventions may be inherent to the intervention, eg, mechanism of action, route of administration, and dosing regimens, but are also likely to be influenced by other study design characteristics such as the model, choice of species, strain, sex, and behavioral outcome measure.

In rodent inflammatory pain models (eg, formalin, complete Freund's adjuvant [CFA], carrageenan, and osteoarthritis), CB₁, CB₂ receptor agonists, and PEA consistently attenuated pain-related behaviors across a range of inflammatory pain models (Table 3). The only exception was for carrageenan-induced inflammation in rats, for which CB₁ receptor agonists did not

significantly attenuate pain-related behaviours. The efficacy of FAAH inhibitors was mixed; pain-related behaviors were significantly attenuated in formalin and CFA but not in osteoarthritis rodent models. In carrageenan models, a species difference was detected in which FAAH inhibitors significantly attenuated pain-related behaviors in mice but not rats. THC also significantly attenuated pain-related behaviors in formalin, CFA, and carrageenan models. Like FAAH inhibitors, the efficacy of CBD was mixed; pain-related behaviors were significantly attenuated in formalin and CFA but not in carrageenan and osteoarthritis rodent models.

In rodent neuropathic pain models, eg, nerve injury, chemotherapy-induced peripheral neuropathy, and diabetes, CB₁ and CB₂ receptor agonists, FAAH inhibitors, and CBD consistently demonstrated antinociceptive efficacy (Table 4). PEA also significantly attenuated pain-related behaviors in nerve injury and chemotherapy models. THC significantly attenuated pain-related behaviors in nerve injury models.

Our companion systematic review and meta-analysis suggests that the most frequently assessed cannabinoids, CB₁ and CB₂ receptor agonists and nonselective agonists, consistently attenuated pain-associated behaviors in a broad range of inflammatory and neuropathic pain models.²¹⁵ Although tested in fewer model types, this was similarly evident for THC. CBD and FAAH inhibitors were not as effective in inflammatory pain models but may be viable candidates for the treatment of neuropathic pain. However, this analysis does not take into account potential side effects such as motor impairment, hypothermia, or anxiolysis that could influence the behavioral outcomes. Prospective preclinical trials are required to better ascertain what factors are influencing the differences in observed efficacy between the different drug classes and model types.

Our companion systematic review and meta-analysis²¹⁵ highlights several differences between the preclinical and clinical assessment of candidate treatments. The number and profile of the drugs assessed preclinically (171 different interventions) differs markedly from those investigated in clinical trials (11 different interventions)⁷⁹ where the predominantly evaluated clinical interventions are pharmacologically complex cannabis-based medical extracts, THC, or THC analogues. Indeed, an unusual feature of the cannabinoid field is that the initiation of clinical trials has not generally followed (or has been very slow to follow) the publication of preclinical data providing evidence of benefit for small-molecule drugs, whereas this is the case for many other drug classes. Moreover, certain indications assessed in clinical trials (eg, pain due to third molar extraction) are not represented in the preclinical literature. The routes of administration also differ, and accompanying pharmacokinetic investigation was only evident in 7% of the studies. However, 69% of studies did confirm CB₁ or CB₂ receptor involvement through the use of antagonists, transgenic mice, or radioligand binding.

Our companion systematic review and meta-analysis²¹⁵ also highlights several weaknesses in face and construct validity of the animal models and there are several study characteristics that can impact pain-associated behavioral outcome measures and the assessment of novel antinociceptive efficacy. The animal cohorts are genetically very similar, converse to the widely heterogeneous patient population. In rats, most studies used either Sprague-Dawley or Wistar strains, although larger effect sizes were evident in the small number of studies reported with Lewis rats. There was a major bias to the use of male rats (91%). Similar findings were found in mice with 81% of studies reporting the use of males across 29 strains. This does not reflect the clinical situation where women are overrepresented among

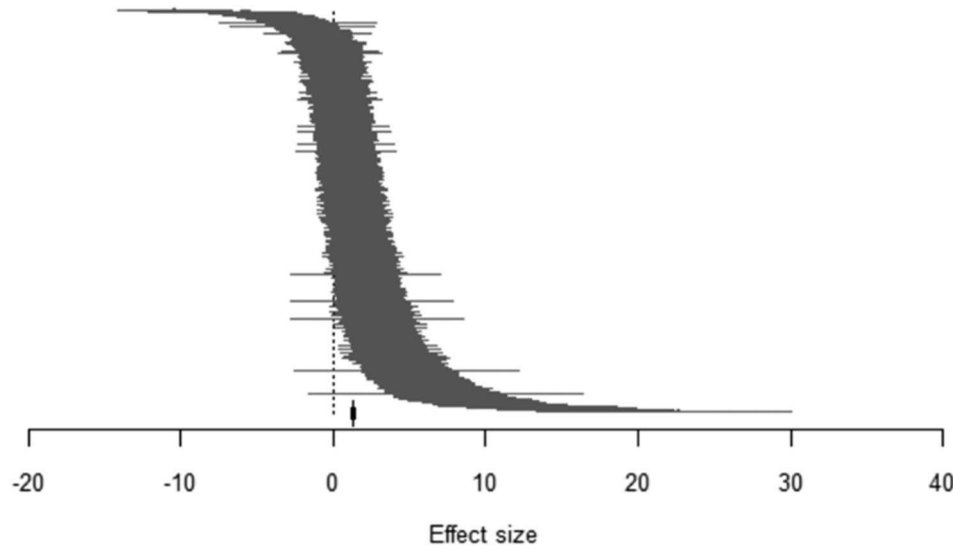


Figure 3. The caterpillar plot of 1544 nested comparisons extracted from 374 studies included in the meta-analysis. A Hedges' G standardised mean difference effect size was calculated for each comparison. Overall effect size = 1.32.²¹⁵

patients with chronic pain.¹⁶⁴ In clinical trials, male and female patients are more equally represented. For example, in a recent systematic review of 36 randomised controlled trials assessing cannabinoids, cannabis, and CBMs, female patients outnumbered (n = 3691) male patients (n = 3613).⁷⁹

More generally, the animal models do not effectively simulate multidimensional clinical pain conditions including the psychological component. Disease or injury is frequently induced in young, otherwise healthy animals contrasting with the clinical situation in which disease or injury predominantly occurs in older patients with comorbidities. The duration of animal studies is usually brief (up to a few weeks), which results in candidate treatments being tested in the early stages of disease onset, which does not adequately reproduce the impact of prolonged clinical pain or address the clinical need for treatment in the later stages of disease. There is an overreliance on evoked limb withdrawal, a measure of hypersensitivity, which is not appropriate for pain that is characterized by sensory loss or spontaneous pain. Careful consideration should be given to the choice of species, strain, sex, and age in relation to the clinical condition being modelled. To limit threats to external validity, researchers

should balance the sexes.³⁹ A broader range of outcome measures that are of clinical relevance and include more complex, ethologically relevant behaviors is required.²⁰² Multi-center testing will increase environmental heterogeneity and study samples thereby improving the generalizability of preclinical findings.²³¹

As with the majority of current preclinical neuroscience and pharmacological research using animal models, the risk of bias is uncertain, due to a generic poverty of reporting sufficient details of the experimental design, conduct, and analysis factors, which govern the veracity of experimental internal validity. There is evidence to suggest that low prevalence of the reporting of measures to mitigate bias tend to give higher estimates of treatment effects.^{48,105} Our meta-analysis did not show a consistent relationship between the reporting of methodological quality criteria and smaller effect sizes; however, larger effect sizes were reported for studies that did not report allocation concealment and sample size calculations.²¹⁵ The methods by which biases were mitigated were also infrequently reported. On the rare occasions where they were reported, the methods were often invalid,²³⁰ eg, randomization by “picking animals randomly

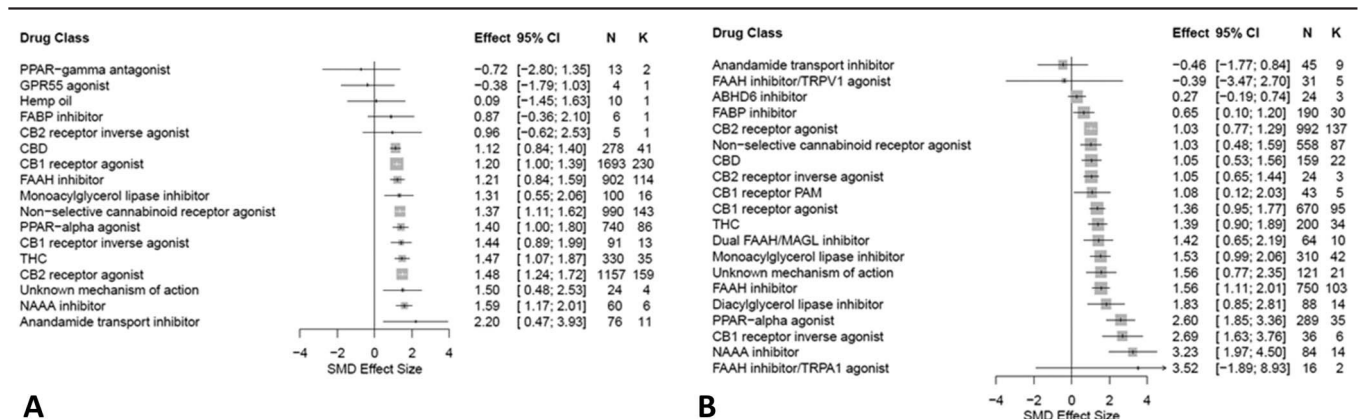


Figure 4. Forest plot of drug classes assessed for antinociceptive efficacy in rat (A) and mouse (B) models of injury-related or pathological persistent pain. The size of the squares represents the weight (%) and its influence on the pooled result. N denotes the number of animals and K the number of comparisons of each subgroup.²¹⁵

Table 3

Summary of the effects of cannabinoids and endocannabinoid system modulators on pain-related behaviour in the most frequently used rodent models of inflammatory pain.

Drug Class	Formalin				Complete Freund's Adjuvant				Carrageenan				Osteoarthritis			
	K	N	Effect size	95 % CI	K	N	Effect size	95 % CI	K	N	Effect size	95 % CI	K	N	Effect size	95 % CI
Anandamide transport inhibitor	10	64	1.97	0.608-3.342	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
CB ₁ receptor agonist	59	496	1.12	0.731-1.502	21	144	0.74	0.378-1.107	ns	ns	ns	ns	3	16	1.22	0.378-1.107
CB ₁ receptor inverse agonist	1	7	1.71	0.427-3.002	NT	NT	NT	NT	5	30	0.44	0.055-0.822	NT	NT	NT	NT
CB ₁ receptor PAM	NT	NT	NT	NT	NT	NT	NT	NT	1	8	2.45	1.067-3.837	NT	NT	NT	NT
CB ₂ receptor agonist	33	265	0.81	0.316-1.307	44	338	1.10	0.829-1.366	10	74	1.39	0.885-1.896	24	173	1.42	0.829-1.366
CB ₂ receptor inverse agonist	3	22	1.05	0.653-1.444	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
CBD	2	16	1.86	1.548-2.169	5	20	1.59	0.71-2.469	ns	ns	ns	ns	ns	ns	ns	ns
Dual FAAH/MGL inhibitor	NT	NT	NT	NT	ns	ns	ns	ns	4	36	1.42	0.25-2.6	NT	NT	NT	NT
FAAH inhibitor	23	184	1.50	1.134-1.862	24	144	1.24	0.702-1.781	ns	ns	ns	ns	ns	ns	ns	ns
FABP inhibitor	ns	ns	ns	ns	15	98	0.78	0.194-1.373	6	36	1.42	0.947-1.895	NT	NT	NT	NT
Monoacylglycerol lipase inhibitor	8	69	2.28	1.382-3.175	ns	ns	ns	ns	8	60	2.67	0.967-4.378	ns	ns	ns	ns
NAAA inhibitor	NT	NT	NT	NT	3	30	1.63	1.235-2.025	12	72	1.91	0.923-2.897	NT	NT	NT	NT
Nonselective cannabinoid receptor agonist	38	253	1.56	1.019-2.11	ns	ns	ns	ns	45	311	0.74	0.202-1.268	ns	ns	ns	ns
PPAR-alpha agonist	21	143	1.73	0.701-2.761	2	16	2.53	0.851-4.205	26	173	3.26	1.51-5.006	9	75	0.98	0.851-4.205
THC	20	118	1.16	0.679-1.643	16	109	2.34	1.687-2.986	5	37	2.13	0.138-4.12	NT	NT	NT	NT

K denotes the number of comparisons and N denotes the number of animals within each subgroup. All entries with an effect size value were statistically significant.

CAB, cannabidiol; CB₁, cannabinoid type 1; CB₂, cannabinoid type 2; CI, confidence interval; FAAH, fatty acid amide hydrolase; FABP, fatty acid-binding protein; MGL, monoacylglycerol lipase; NAAA, N-acyl ethanolamine-hydrolyzing acid amidase; NT, not tested; ns, not significant; PAM, positive allosteric modulator; PPAR, peroxisome proliferator-activated receptor; THC, tetrahydrocannabinol.

Adapted from the study by Soliman et al.²¹⁵

from a cage" rather than computer-generated random sequence, or determining sample size based upon reported sample size norms rather than a power calculation. Thus, there is a critical need for transparency of reporting all experimental details so that the quality of research can be assessed and the validity of the outcomes inferred.

The analysis did not suggest evidence of publication bias. It is also to be expected that animal studies will yield larger effect sizes in comparison to clinical studies due to the more homogenous nature of the laboratory animal study population and better opportunity to control experimental variables, reducing the observed variance and making direct comparison of effect sizes between animal and human studies impossible.

Our accompanying systematic review and meta-analysis²¹⁵ therefore highlights the need for improvements in experimental design and, perhaps even more importantly, the reporting of experimental design and analysis features in sufficient detail such that primary research can be reproduced and meta-analysed. Notwithstanding the caveats relating to animal studies of analgesia, substantial evidence from animal model experiments supports the hypothesis of cannabinoid-induced analgesia in inflammatory and neuropathy conditions.

6. Side effects relevant to preclinical pain studies

An important consideration in the evaluation of any pain medication is whether analgesic efficacy is accompanied by adverse side effects that limit therapeutic potential, or additional beneficial effects that might enhance therapeutic potential. The present evaluation of on-target "side effects" is restricted to effects of cannabinoids/endocannabinoid system modulators administered to rodents in adulthood. Additional preclinical research is needed to specifically assess potential harms in vulnerable populations (eg, older adults, and the developing fetal and adolescent brain^{40,41,91}). In this section, we consider both potential harms and benefits of pharmacological strategies assessed *specifically* in animal pain models, particularly as they may emerge with chronic dosing. We also consider pharmacological effects (eg, motor impairment, hypothermia, or anxiolysis) that could influence interpretation of pain-related behavior. Animal models have been used to investigate an array of mechanisms and sites of analgesic action using a much broader array of mechanistically distinct compounds compared with those evaluated in the clinical literature. Most therapeutic interventions that show promise in preclinical studies,

Table 4

Summary of the effects of cannabinoids and endocannabinoid system modulators on pain-related behaviour in the most frequently used rodent models of neuropathic pain.

Drug Class	Nerve injury				Chemotherapy				Diabetes			
	K	N	Effect size	95 % CI	K	N	Effect size	95 % CI	K	N	Effect size	95 % CI
Anandamide transport inhibitor	1	5	7.95	3.26-12.643	NT	NT	NT	NT	NT	NT	NT	NT
CB ₁ receptor agonist	102	756	1.17	0.842-1.5	31	173	1.47	1.007-1.937	30	183	1.74	1.234-2.248
CB ₁ receptor inverse agonist	4	36	1.71	1.171-2.248	NT	NT	NT	NT	2	12	4.53	1.932-7.127
CB ₂ receptor agonist	87	639	1.15	0.812-1.497	27	192	1.55	1.117-1.983	10	88	1.29	0.772-1.818
CBD	9	72	1.86	1.307-2.406	10	76	1.31	0.594-2.021	7	49	1.08	0.257-1.896
Diacylglycerol lipase inhibitor	1	6	3.69	1.235-6.148	ns	ns	ns	ns	NT	NT	NT	NT
Dual FAAH/MGL inhibitor	2	12	2.08	1.348-2.808	NT	NT	NT	NT	NT	NT	NT	NT
FAAH inhibitor	59	439	1.75	1.212-2.281	19	148	1.96	1.048-2.868	17	180	2.53	1.455-3.604
Monoacylglycerol lipase inhibitor	16	119	1.31	0.774-1.84	12	80	1.42	0.179-2.653	NT	NT	NT	NT
NAAA inhibitor	5	34	4.54	2.594-6.496	NT	NT	NT	NT	NT	NT	NT	NT
Nonselective cannabinoid receptor agonist	75	504	1.18	0.825-1.536	ns	ns	ns	ns	8	48	1.89	0.335-3.437
PPAR-alpha agonist	26	229	1.14	0.493-1.786	11	74	2.54	1.341-3.746	NT	NT	NT	NT
THC	9	86	1.26	0.838-1.677	NT	NT	NT	NT	NT	NT	NT	NT

Adapted from the study by Soliman et al.²¹⁵

K denotes the number of comparisons and N denotes the number of animals within each subgroup. All entries with an effect size value were statistically significant.

CAB, cannabidiol; CB₁, cannabinoid type 1; CB₂, cannabinoid type 2; CI, confidence interval; FAAH, fatty acid amide hydrolase; FABP, fatty acid-binding protein; MGL, monoacylglycerol lipase; NAAA, N-acyl ethanolamine-hydrolyzing acid amidase; NT, not tested; ns, not significant; PPAR, peroxisome proliferator-activated receptor; THC, tetrahydrocannabinol.

nonetheless, fail in clinical trials, albeit for different reasons (ie, efficacy, side-effect profile, lack of adequate target engagement, therapeutic indication, variability in clinical populations, and clinical primary outcome measure); this failure rate is not unique to cannabinoids. Moreover, animal models may not adequately capture adverse side effects that may be problematic in humans, and adverse side effects are often not systematically studied in animal experiments. Nevertheless, preclinical studies have provided evidence of both beneficial (ie, antinociception, antiemetic, antispasticity, antistress, and anti-anxiety-like effects) and adverse (ie, reward or aversion, dependence, tolerance, and motor and memory impairment) effects of cannabinoids and endocannabinoid system modulators in otherwise normal animals (for review, see Refs. 183,184). Here, we primarily consider “on-target” pharmacological effects of cannabinoids in pain models only, including assessments of cannabimimetic effects (ie, in the classic cannabinoid tetrad), tolerance, physical dependence, reward/reinforcement, opioid-sparing effects, antinociceptive synergy, and effects on stress-, anxiety-, and depression-related behavior in animal models of pain.

6.1. Cardinal signs of cannabinoid type 1 receptor activation

The classic cannabinoid tetrad assesses cardinal signs of CB₁ activation (ie, hypoactivity, reduction in body temperature, catalepsy, and tail-flick antinociception)^{156,214} in rodents and is produced by all CNS-penetrant CB₁ agonists, including THC. These cannabimimetic effects are consistent with localisation of CB₁ to motor and limbic regions in rodent brain.¹⁰⁴ Drug-induced motor impairment can mask detection of antinociception in rodents. Consequently, preclinical studies must show that antinociceptive effects of cannabinoids observed in behavioral studies are not artifacts of motor impairment. Although motor effects complicate behavioral evaluation of antinociceptive effects of cannabinoids, they do not preclude the existence of antinociceptive mechanisms. Our accompanying systematic

review revealed that a minority (33%) of studies assessed the effects of the drugs on motor activity.²¹⁵ Direct CB₁ agonists and nonselective cannabinoid receptor agonists that penetrate the CNS have potential to produce undesirable CB₁-mediated pharmacological effects in humans (eg, psychoactivity, and motor and memory impairment) (for review, see Refs. 183,184). Electrophysiological measures of activity of nociceptive neurons and/or neurochemical measures of noxious stimulus-evoked neuronal activation provide independent lines of evidence that nonselective cannabinoid receptor agonists, CB₁ agonists, FAAH inhibitors, and CB₂ agonists suppress nociceptive processing in rodents (for review, see Refs. 98, 99, 217, 240). Nonselective cannabinoid receptor agonists (eg, WIN55,212-2) suppress electrophysiological^{112,114,115,158} and neurochemical markers of pain-evoked neuronal activation²²⁶ as well as pain behavior at doses that do not alter body temperature or produce immobility.^{186,199} Drug-induced reductions in body temperature cannot explain electrophysiological or behavioral indicators of antinociceptive efficacy.^{158,170,228} WIN55,212-2-induced reduction of evoked hypersensitivity in the chronic constriction injury (CCI) model of neuropathic pain, and side effects (motor incoordination, catalepsy, and sedation), can occur at similar ED₅₀s in mice³; however, antinociceptive effects of WIN55,212-2 correlate with suppression of firing in nociceptive neurons and is reported to outlast these motor effects.¹⁵⁸

Drug development efforts have focused on elucidating therapeutic potential of small molecules that engage targets within the endocannabinoid system that lack unwanted cannabimimetic effects associated with direct CB₁ activation (for review, see Refs. 50, 113, and 139). Cannabinoid type 2 agonists, FAAH inhibitors, MGL inhibitors (at appropriate doses), CB₁-positive allosteric modulators (PAMs), peripherally restricted nonselective cannabinoid receptor agonists (PrNM1¹⁶⁶) have all been shown to suppress pain behavior without unwanted motor effects of CB₁ agonists (for review, see Refs. 98, 99, 217, and 240). High-dose MGL inhibitors can elicit tetrad effects, whereas

a dual FAAH/MGL inhibitor JZL195 suppressed neuropathic nociception in the mouse CCI model with an ED50 4 times lower than that which produced side effects (motor incoordination, catalepsy, and sedation),³ although brain-permeant and -impermeant FAAH inhibitors lack such effects.^{5,73,197,223} Dual FAAH/MGL inhibitors represent a pharmacological strategy to elevate both AEA and 2-AG and produce antinociceptive efficacy without producing unwanted cannabimimetic effects in the tetrad.⁶⁶ Cannabinoid type 1 knockout mice have also been used to assess the impact of CB₂ activation without the confound of CB₁-mediated side effects.^{56,66,126,208} Such studies show that mixed cannabinoid agonists and CB₂ agonists can engage CB₂ receptors independently of CB₁ to alleviate neuropathic pain-related behavior induced by paclitaxel administration or spinal nerve ligation.^{56,126}

The abundance of CB₁ in brain regions controlling motor activity and memory accounts for adverse side effects of mixed and CB₁-preferring strategies.¹⁰⁴ Very few studies evaluate possible memory impairment induced by pharmacological treatments in laboratory animal pain models. However, studies that involve learning approaches document that rodents in pathological pain states can show conditioned place preferences^{50,84} or perform operant responses^{102,166} to chambers/tasks associated with pain relief; memory impairment would preclude demonstrations of efficacy in such studies. Peripherally restricted CB₁ agonists, CB₂ agonists, MGL inhibitors, and CBD represent cannabinoid modulators that have shown efficacy in such assays in animal pain models.^{50,84,166}

6.2. Tolerance

Tolerance, the loss of therapeutic efficacy with repeated administration, is undesirable in an analgesic and can lead to dose escalation and potential for misuse and abuse. In animal models of pathological pain, tolerance to therapeutic efficacy develops to direct-acting CB₁ agonists, nonselective cannabinoid receptor agonists, as well as high, but not low, doses of MGL inhibitors,^{56,142,212} presumably via downregulation and desensitization of CB₁ receptors. In a mouse model of chemotherapy-induced neuropathic pain induced by paclitaxel, tolerance developed to the antinociceptive effects of THC and other nonselective cannabinoid receptor agonists (CP55,940 and WIN55,212-2), as well as to other classic cannabimimetic effects.^{56,212} Low-dose chronic infusion of WIN55,212-2 and AM1241 (CB₂ agonist) were associated with sustained antinociceptive efficacy in the paclitaxel model without motor impairment.¹⁹⁹ Tolerance develops more quickly to high compared with low doses of the centrally acting CB₁-preferring ligands.^{56,212} By contrast, antinociceptive tolerance is typically absent in neuropathic as well as inflammatory pain models after repeated administration of CB₂ agonists,^{56,199,212} brain-permeant and -impermeant FAAH inhibitors,^{38,212,213} low-dose (but not high-dose) MGL inhibitors,¹⁴² CB₁ PAMs,^{128,212} and a peripherally restricted CB₁ agonist.¹⁶⁶ The FAAH/MGL dual inhibitor JZL195 exhibited greater efficacy than FAAH or MGL inhibitor alone and did not produce tolerance, exhibiting an improved therapeutic window compared with direct cannabinoid CB₁ agonists.³ Further preclinical studies are necessary to determine whether different therapeutic strategies (ie, biased CB₁ agonism, peripherally restricted nonselective cannabinoid receptor agonists) can separate therapeutic effects from unwanted side effects.

6.3. Physical dependence

Receptor antagonists have been used to precipitate a withdrawal syndrome, a sign of physical dependence, in mice subjected to

pathological pain states. In mice rendered neuropathic with paclitaxel, the CB₁ antagonist rimonabant precipitates signs of physical dependence (ie, paw tremors) in mice treated chronically with orthosteric cannabinoid agonists (ie, THC, CP55940, and WIN55,212-2); severity of withdrawal symptoms was dose-related.^{56,212} In the same neuropathic pain model, both tolerance and rimonabant-precipitated withdrawal signs were produced by the MGL inhibitor JZL184 but not by a CB₂ agonist,^{56,212} brain-permeant or -impermeant inhibitors of FAAH,²¹³ or a CB₁ PAM.²¹² Where present, physical dependence was induced by challenge with CB₁, but not CB₂, antagonists and in animals receiving direct-acting agonists (or high-dose MGL inhibitor) that penetrated the CNS. By contrast, neither CB₁ PAMs nor brain-permeant or -impermeant inhibitors of FAAH were associated with signs of physical dependence after 3 weeks of once daily administration in paclitaxel-treated mice.^{212,213} Notably, CB₁ antagonist-precipitated withdrawal syndromes (ie, paw tremors) lacked the more striking somatic (ie, jumping) and autonomic (ie, diarrhea) signs associated with naloxone-precipitated opioid withdrawal.^{9,212}

6.4. Reward/reinforcement

Tetrahydrocannabinol can produce both rewarding and aversive effects in laboratory animals.^{35,151,216} Rewarding properties of cannabinoids have historically been demonstrated in otherwise healthy, young, male rodents, which may not necessarily mimic the situation in people with chronic pain. In pathological pain states, interpretation of positive reinforcing effects of cannabinoids (ie, reward in the typical drug abuse sense) should be differentiated from negative reinforcing effects (ie, removal of an aversive pain state). Preclinical research involving rodent pain models has investigated therapeutic strategies targeting the endocannabinoid system that hold promise for suppressing pain without producing abuse liability. Using a classic drug self-administration approach, rats rendered neuropathic by a spared nerve injury self-administered a CB₂ agonist in a CB₂ receptor-dependent manner; naive animals did not reliably self-administer the drug.¹⁰² Moreover, CB₂ agonists did not produce CPP.^{93,130} These observations suggest that the CB₂ agonists were not inherently reinforcing in the absence of the pathological pain state. In the absence of pathological pain, CB₁ PAMs ZCZ011 and GAT211 do not produce CPP when administered alone.^{129,212} Moreover, ZCZ011 did not substitute for CB₁ agonists CP55940 or AEA in a drug discrimination assay,¹²⁹ suggesting that it does not produce cannabimimetic side effects. Inhibition of MGL with MJN110 also reversed a negative affective state associated with paclitaxel treatment using a CPP approach.⁵⁰ Similarly, a nonrewarding dose of CBD produced reward-related effects in the presence of pain due to incisional injury.⁸⁴ Thus, in the absence of a pathological pain state, CB₁ PAMs and endocannabinoid deactivation inhibitors did not produce rewarding or aversive effects when administered alone in rodents. These studies raise the possibility that components of the endocannabinoid signaling system may be targeted for therapeutic benefit without the rewarding properties of direct CB₁ activation in the CNS.

6.5. Opioid-sparing effects and antinociceptive synergy

Opioids remain a mainstay of pain management but also produce tolerance, physical dependence, reward, constipation, and respiratory depression, among other effects. Efficacy of adjunctive therapies for suppressing pathological pain and producing

opioid-sparing effects has, consequently, been evaluated. Tetrahydrocannabinol produced synergistic antinociceptive effects with morphine in both arthritic (Freund's adjuvant-induced) and naive rats⁴⁵; additive, rather than synergistic, interactions were reported for unwanted side effects (eg, tetrad and motor ataxia).⁴⁵ WIN55,212-2 produced synergistic antinociceptive interactions with morphine in mice with CCI of the sciatic nerve but only additive effects on motor coordination.³ Brain-permeant and -impermeant inhibitors of FAAH,²¹² MGL inhibitors,²³⁷ dual FAAH-MGL inhibitor,⁶⁶ CB₂ agonists,^{93,130} and CB₁ PAMs²¹¹ produce synergistic antinociceptive effects with morphine in neuropathic pain models. The CB₂ agonist JWH133 produced additive antinociceptive effects with morphine in the formalin test.²⁴⁵ Notably, in a mouse neuropathic pain model, brain-permeant (URB597) and -impermeant (URB937) FAAH inhibitors,²¹³ a CB₁ PAM (GAT211,²¹¹), and CB₂ agonists (AM1710, LY2828360;^{9,153,212}) suppressed development of morphine tolerance without enhancing naloxone-precipitated opioid withdrawal. Multiple CB₂ agonists attenuated opioid tolerance and naloxone-precipitated opioid withdrawal in neuropathic mice.^{9,130,153,155,212} Cannabinoid type 2 agonists produce synergistic antinociceptive effects with opioids and also attenuate opioid-induced respiratory depression.^{236,246} Neither FAAH inhibitors (URB597 and URB937) nor a CB₁ PAM (GAT211) enhanced naloxone-precipitated opioid withdrawal in paclitaxel-treated mice.^{212,213} Evaluations of other opioid side effects in the same studies (ie, slowing of GI motility and reward) have typically used normal animals not subjected to pathological pain states. Nonetheless, lowering opioid doses required to elicit therapeutic effects could enhance therapeutic ratios. Most studies have combined opioids with endocannabinoid modulators that themselves lack observable cannabimimetic side effects (JWH015, MJN110, URB597, and URB937); consequently, additivity of adverse side effects would not be expected. Interestingly, CB₂ agonists (JWH015 and LY2828360) produced synergistic anti-allodynic effects with morphine in models of inflammatory (formalin), postoperative (paw incision), and neuropathic (SNL) nociception but did not produce synergy for nociceptive pain.^{93,130} Synergy between CBD and morphine has also been reported in the acetic acid-induced writhing model,¹⁷⁶ and coadministration of THC and morphine reduced the second phase of formalin-evoked nociceptive behaviour in rats to a greater extent than either drug alone.⁷⁸ More work is necessary to examine whether cannabinoids alter other unwanted side effects of opioids (eg, slowing of GI motility) in pain models and evaluate the clinical relevance of these findings

6.6. Synergism of antinociceptive efficacy: nonopioid analgesics

Tetrahydrocannabinol has been shown to produce synergistic antinociceptive interactions with gabapentin in the mouse CCI model of neuropathic pain.⁹ Synergy between CBD and THC, and between PEA and gabapentin, is also reported in chemotherapy-induced neuropathic pain models,^{66,67,141} between CBD and THC in the mouse CCI model of neuropathic pain,³⁴ and between PEA and acetaminophen (paracetamol) in the rat streptozotocin-induced model of diabetic neuropathy.⁵³ The COX-2 inhibitor celecoxib increased the antihypersensitivity activity of the CB₁ agonist Met-F-AEA and the CB₂ agonist AM1241 in the streptozotocin model.²⁶ Low doses of the MGL inhibitor JZL184 and the nonselective COX inhibitor diclofenac synergistically attenuated mechanical allodynia and additively reduced cold allodynia in the mouse CCI model of neuropathic

pain.⁴⁹ Furthermore, coadministration of the FAAH inhibitor URB597 and diclofenac yielded synergistic antinociceptive effects in the acetic acid-induced abdominal stretching model of visceral nociception in mice.¹⁷⁴ In other work, AEA coadministered with either ibuprofen (nonselective COX inhibitor) or rofecoxib (selective COX-2 inhibitor) resulted in synergistic antinociceptive effects in the rat formalin test.^{97,101} Mechanistically, it is important to note that there are a number of interactions between COX/NSAIDs/acetaminophen and cannabinoids/endocannabinoid system, including inhibitory effects of NSAIDs or an acetaminophen metabolite on endocannabinoid catabolism or transport (for review, see Ref. 189). Thus, adjunctive therapies could enhance therapeutic ratios of existing treatments for both inflammatory and neuropathic pain. Cannabinoid type 1 PAMs enhance antinociceptive and unwanted side effects of orthosteric CB₁ agonists^{128,212} and produce synergistic antiallodynic effects with FAAH and MGL inhibitors without enhancing other tetrad parameters.²¹²

6.7. Beneficial on-target pharmacological effects

Beneficial on-target pharmacological effects (ie, in suppressing stress, anxiety, nausea, and producing improvements in sleep) of cannabinoids (for review, see Refs. 183, and 184) could contribute to perceived therapeutic benefits in relevant pathological pain states. However, only small numbers of studies have evaluated such features specifically in animal pain models. Indeed, our accompanying systematic review revealed that only 3% of studies assessed the anxiolytic or antidepressant-like effects of the drugs in the animal models of injury-related or pathological persistent pain.²¹⁵ Systematic preclinical studies are necessary to determine whether, rather than purely modulating sensory thresholds, cannabinoid-based modulation of affective behavior could contribute to therapeutic efficacy of analgesics by attenuating symptoms that exacerbate pain.

6.8. Stress, anxiety, depression, and pain-depressed behavior

The therapeutic potential of cannabinoids/endocannabinoid system modulators has been assessed in animal models of anxiety, stress, and depression (for review, see Refs. 76, 90, 165, 183, 184, and 187). However, fewer studies have evaluated such therapeutic effects within the context of pathological pain states (for review, see Refs. 42, 80, 217, and 240). The CB₂ agonist GW405833 reduced immobility (ie, a measure of depression-like behavior) in the forced swim test and allodynia (although mediation by CB₂ was not assessed), whereas the antidepressant desipramine primarily attenuated immobility time.¹²⁰ In the mouse monosodium iodoacetate model of osteoarthritis pain, the CB₁ agonist ACEA and the CB₂ agonist JWH133 ameliorated nociceptive and affective alterations, and ACEA also improved associated memory impairment.¹⁴⁷ In rats with CCI, the brain-permeant FAAH inhibitor URB597 but not the brain-impermeant FAAH inhibitor URB937 suppressed immobility in the forced swim test, and attenuated novelty-induced suppression of feeding and CCI-induced reductions in hippocampal neurogenesis, all indicative of antidepressant-like effects, whereas both URB597 and URB937 suppressed allodynia.¹³² Thus, the CNS-penetrant FAAH inhibitor produced beneficial effects on both evoked pain and pain-induced depression-like behavior. Rats with spared nerve injury exhibited an anxiety-like phenotype relative to shams; repeated CBD prevented anxiety-like behavior in the open field test, elevated plus maze and novelty-induced suppression of

feeding test, in addition to suppressing allodynia via a 5-HT_{1A} receptor mechanism.⁵² Thus, effects of CBD revealed in that study are likely to be independent of cannabinoid receptor activation.

In addition to evaluations of responses to evoked pain (pain-stimulated behaviors), an emerging preclinical literature has evaluated pain-depressed behavior (eg, marble burying, nestlet shredding, and intracranial self-stimulation thresholds). Although the extent to which assays of pain-depressed behavior assess stress-, anxiety-, or depression-related behavior is uncertain, they represent an attempt to more completely model the multifaceted complexities associated with pathological pain states.¹⁷⁷ Assessing the effects of drugs on pain-depressed behavior may also help to distinguish between antinociception and motoric side effects of putative analgesics. Tetrahydrocannabinol and CP55,940 produced antinociception for pain-stimulated responding in an acid-induced writhing assay but exacerbated pain-depressed behavior (ie, noxious stimulus-induced suppression of intracranial self-stimulation thresholds).^{146,150} By contrast, the FAAH inhibitor URB597 suppressed both pain-stimulated as well as pain-depressed behavior.¹⁴⁵ An MGL inhibitor (MJN110), CB₂ agonist (LEI101), nonselective cannabinoid receptor agonist (CP55940), and reference analgesics (morphine, gabapentin, and valdecoxib) all reversed pain-stimulated behavior (ie, mechanical hypersensitivity) as well as pain-depressed behavior (ie, deficits in marble burying) in mice with CCI.⁵⁰ By contrast, the benzodiazepine diazepam reversed neither dependent measure, whereas a kappa-opioid receptor agonist (U69593) and an FAAH inhibitor (PF3845) reversed deficits in mechanical hypersensitivity only.⁵⁰ Deficits in marble burying resolved within a week of surgery,⁵⁰ and not all neuropathic pain models are associated with deficits in marble burying/nestlet shredding.²¹² Thus, the translational relevance of these behaviors to pain and/or anxiety, consequently, remain unclear but may they provide insights of some relevance to general health/quality of life.

6.9. Sleep

Effects of cannabinoids on sleep have not been assessed in animal pain models. Clinical data on the FAAH inhibitor PF-04457845 in cannabis use disorder have suggested improvements in both withdrawal and sleep.⁵¹ More work is necessary to ascertain presence of sleep disruptions in laboratory animal models of pathological pain states and ascertain whether endocannabinoid modulators could restore such deficits.

7. Pharmacokinetics of cannabinoids

In addition to the mechanisms of drug action that affect the endocannabinoid system, and cannabinoid structure-activity relationships (ie, pharmacodynamics), the assessment of drug pharmacokinetics is of critical importance both for better understanding the preclinical pharmacology, and for improving animal-to-human translation.²³⁵ The pharmacokinetic processes of drug absorption to the systemic circulation, its distribution to tissues of interest, as well as its metabolism and excretion will determine how much drug, after a particular dose delivered via a particular route, will reach its site of action. In addition, drug pharmacokinetics will determine the duration of a drug's effect, and additional parameters related to drug–drug interactions and dose adjustments required in cases when organs such as liver or kidneys do not function properly.

Animal models provide advantages for investigating drug pharmacology because endogenous systems often cannot be manipulated safely in humans. However, because drug pharmacokinetics can be investigated directly in humans, this section of the article will also discuss clinically relevant pharmacokinetic data, where possible. Pharmacokinetic properties of cannabinoids, particularly their absorption, demonstrate substantial variability as a function of both route of administration and the specific formulation in which the cannabinoids are delivered. Human studies repeatedly demonstrate large intersubject variability in the pharmacokinetics of identical cannabinoid products, the basis of which is currently not well understood. Therefore, comparisons of nonhuman and human pharmacokinetics can have important translational relevance.

7.1. Absorption

Cannabinoid absorption is generally faster via the inhalational route than oral route, resulting in a more rapid onset of pharmacological effects, and shorter time to peak effect. In addition, cannabinoid delivery via the inhalational route circumvents the variability in oral absorption processes that are due to first pass metabolism. The limitations of inhalational routes include the variability in interindividual efficiency that is caused by differences in inhalation techniques, respiratory tract irritation during inhalation, and the inconvenience or lack of adherence associated with smoked, vaporised, or nebulised cannabinoid products. Although most preclinical experiments are performed with injected or oral/gastric administration of cannabinoids, most clinical therapeutic studies have used either inhaled products, oromucosal products such as nabiximols/Sativex, or oral THC and its analogues.

The reported oral bioavailability of cannabinoids, and THC in particular, varies as a function of drug vehicle and the coingested food. For example, orally ingested cannabis-containing cookies demonstrate only 6% THC bioavailability, whereas THC dissolved in sesame oil delivered in soft gelatin capsules is ~20% bioavailable (for review, see Ref. 122). A similar pattern of improved oral bioavailability approaching 30%, when dissolved in sesame oil, has been reported in rat studies with both THC and CBD.¹³¹

The bioavailability of THC delivered via smoked cannabis products reportedly varies between 18% and 50%.¹⁸⁰ In a small study comparing absorption of a physiologically compatible, nebulised inhalation solution of THC with that of intravenous THC in 8 healthy volunteers, the average bioavailability of THC delivered via the inhalational route was approximately 28%,¹⁷² somewhat comparable to average bioavailability with smoking. However, the measured area under the THC concentration–time curve (AUC) among study participants varied more than 5-fold after intravenous administration, and more than 15-fold after inhalation. Adverse effects, as expected, were more prominent with intravenous dosing.¹⁷²

After oral THC administration, the time to reach maximum plasma concentration (T_{max}) has been reported between 30 and 120 minutes (but up to 6 hours in some studies) and is comparable between young and older adults.⁴ Similarly, a wide range of T_{max} values, between 1 and 8 hours, has been reported with rat studies of THC, depending on the fasted/fed status and the formulation in which THC is administered.^{131,198} However, peak plasma concentrations (C_{max}) of THC seem to be highly variable, with 4- to 7-fold differences among individuals, particularly in older adults.⁴ With single oral doses of THC, the average C_{max} ranged between 1.5 and 3 ng/mL after 3 to 6.5 mg

doses,⁴ and approximately 7 ng/mL after 20 mg oral THC (dronabinol) administration.¹⁷¹ With inhaled doses, 0.053 mg/kg nebulised THC (mean dose 3.5 mg)¹⁷² resulted in average C_{max} of 20 ng/mL, and with smoked cannabis, THC doses of 0.25 mg/kg body weight (14–22 mg per cigarette) have resulted in mean C_{max} of 48 ng/mL. The plasma concentration profiles of key THC metabolites, namely 11-COOH-THC- and 11-OH-THC, also vary substantially among people. With such variability in absorption and plasma concentrations of some cannabinoids, it is not surprising that clinical trial results are inconsistent.⁷⁹

Cannabidiol, considered by many as devoid of the psychotropic side effects of other cannabinoids, has garnered attention in recent years. After oral administration, CBD follows close-to-linear absorption (eg, C_{max} of 530 ng/mL with 3000 mg/d, and 780 ng/mL with 6000 mg/d oral dosing), but high-fat meals can increase CBD plasma exposure and peak plasma concentrations by 4- to 5-fold.^{82,221} Following doses of 10 mg CBD with 10 mg THC either in an oral capsule or oromucosal nabiximol spray, mean reported C_{max} was 2.5 to 3 ng/mL for CBD and 6.1 to 6.4 ng/mL for THC.¹³⁸

Cannabinoids are highly lipophilic molecules with low aqueous solubility and are susceptible to degradation and oxidation, especially in a solution. Drug formulation can thus play a crucial role in increasing the solubility and physicochemical stability of cannabinoid drugs, thus improving their pharmacokinetic properties.²⁵ Commonly used strategies in marketed products include pH adjustment, use of cosolvents, the use of micelles and nano-emulsions, complexation with cyclodextrins, or encapsulation in lipid-based formulations such as liposomes and nanoparticles. Even simple approaches such as administration of THC and CBD in lipid-rich oral formulations can increase plasma bioavailability of cannabinoids 2- to 3-fold, compared with lipid-free formulations.²⁴⁸

Formulations that are based on self-(nano)emulsifying drug delivery technology (SEDDS) have been proposed as a means of improving the oral bioavailability of drugs that show poor aqueous solubility, including cannabinoids. This approach results in a consistent increase in oral bioavailability of both THC (9-fold) and CBD (6-fold) in rats,³⁷ and a 1.5-fold and 2.2-fold increase, respectively, in humans.³⁶

Despite structural similarities, oral bioavailability differs between different cannabinoids. Nabilone, a synthetic cannabinoid derivative, is only slightly structurally different from THC (and dronabinol); however, it has substantially higher bioavailability than dronabinol (95% vs 10%–20%). One proposed difference relates to relative affinity to chylomicrons, and therefore the extent of lymphatic absorption of a particular cannabinoid.⁸⁵ Drug formulations containing long-chain triglycerides may substantially improve the lymphatic absorption of selected cannabinoids.⁸⁵ Cannabinoids have demonstrated immunomodulatory effects *in vitro*; however, plasma concentrations achieved with clinically relevant dosing are well below those shown to affect lymphocyte function. By choosing the appropriate lipid-based formulation, lymphatic absorption of orally administered cannabinoids can be improved to achieve 100-fold and 250-fold higher lymph (vs plasma) concentrations of THC and CBD, respectively, which could help in the targeting of conditions where neuroimmunological effects of cannabinoids are of interest.^{247,248}

Although there is high variability with the oral routes, consideration of variability related to smoked or vaporised delivery is also important. In pulmonary delivery routes, the depth of inhalation, how long breath is held for, and vapouriser temperature all affect cannabinoid absorption.²⁵ Inhalational devices that are temperature-regulated to control the inhaled doses may allow less variable pharmacokinetic profile of inhaled cannabinoids.⁷⁰

The transdermal route of administration provides an alternative approach for systemic delivery of lipophilic compounds with highly variable oral absorption, but more research is required to optimise these delivery systems and assess their efficacy and safety profiles.^{188,229}

7.2. Distribution

Most naturally occurring cannabinoid receptor ligands are lipophilic, readily penetrate the blood–brain barrier, and permeate well to lipid-rich tissues such as the brain and peripheral fat. Cannabinoids display slower elimination rates from these tissues compared with plasma.²⁴ Tetrahydrocannabinol achieves higher concentration in rodent brain than in plasma, and CBD achieves approximately a 1:1 ratio.¹⁰⁶ Many of the synthetic ligands behave similarly. For example, WIN 55,212-2 and MK-9470, cannabinoid receptor ligands commonly used in preclinical studies, demonstrate similarly high brain:plasma penetration ratio in rodents.^{163,194}

In rhesus monkeys exposed to either intermittent (days) or long-term (years) cannabis consumption, the pharmacokinetics of single-dose subcutaneous THC were not substantially different.⁸⁷ However, physiological responses of change in core temperature, or behavioral response time were different, and less profound, in monkeys with chronic exposure, likely suggesting physiological/behavioral tolerance to THC. Although blood THC levels were highest at 30-minute postinjection, maximal effects on temperature and response rate did not occur until 120-minute after injection, after blood THC levels had substantially decreased, suggesting a nonlinear relationship between blood concentration of THC and its pharmacodynamic effects.

The existence of peripheral antinociceptive mechanisms^{32,81} (for review, see Refs. 107, 152, 169, 196, 204, 240, and 250) prompted development of peripherally restricted cannabinoid receptor ligands. However, developing such compounds that are not lipophilic but have high binding affinity to CB receptors has been challenging. Several compounds meeting the desired criteria of limited brain penetration with high affinity to CB₁ and CB₂ receptors have been synthesized.² Some achieve 0.16 to 0.18 brain:plasma ratio and produce antinociceptive effects in a rat model of neuropathy without producing catalepsy, but are yet to be evaluated clinically. Brain-impermeant inhibitors of endocannabinoid metabolism have also been developed and show promise in preclinical studies of inflammatory and neuropathic pain.^{38,100,212,213}

There may be sex-specific differences in cannabinoid pharmacokinetics. Plasma levels of THC after intraperitoneal injection are comparable between male and female rats but levels of THC metabolites in brain tissue, including 11-OH-THC, the major active metabolite, were higher in female than in male rats. When behavioral effects such as response to heat nociception and catalepsy were tested, SKF525A, a cytochrome P450 inhibitor, attenuated THC-induced antinociception and catalepsy in female but not in male rats. Greater levels of active THC metabolites produced by females could potentially contribute to greater behavioral effects of THC in female compared with male rats.²²⁵ Human studies have found somewhat similar tendencies, with 11-OH-THC levels higher in females after THC administration, along with higher subjective ratings of drug effects.²¹⁰

7.3. Metabolism and excretion

Cannabinoids undergo a variety of metabolic processes in the gut, the liver, and various other tissues. Many metabolites have been identified, but the relative activity and toxicity of each is

unknown. Once absorbed, THC is primarily oxidized by the cytochrome P450 hepatic mixed-function oxidase system to equipotent 11-hydroxy-THC (11-OH-THC), and further metabolized to inactive 11-COOH-THC. Most studies assessing the metabolic profile of THC have measured the plasma concentrations of these 2 metabolites, 11-OH-THC and 11-COOH-THC. The area under the concentration–time curve (AUC) of the psychoactive metabolite 11-OH-THC is 4 to 6 times lower compared with the AUC of THC itself after inhalational or IV administration, but overall, 11-OH-THC has a similar pharmacokinetic profile and elimination half-life.^{171,172}

11-COOH-THC, the main cannabinoid metabolite detected upon urine drug screens, has average terminal elimination half-life of 3 to 5 days. However, due to high interpatient variability, both in adults and adolescents, 11-COOH-THC can be detected in plasma or urine a month or more after biochemically verified abstinence.²⁰⁹

Cannabidiol undergoes multiphasic elimination. Its effective half-life estimates ranged from 10 to 17 hours in humans, and the terminal elimination half-life is approximately 2 to 3 days.²²¹ The effective half-life of some of its metabolites, including 7-carboxycannabidiol (COOH-CBD), is around 24 hours, with plasma concentrations in hundreds of ng/mL detectable many days after single-dose administration. In rats and mice, the half-life of CBD has been reported in the range of 1 to 4 hours, with some unexplained variability among administration by different routes and in different formulations.^{20,54,137,244}

When oral CBD (Epidiolex) was administered in people with variable extent of liver impairment, the total exposure (AUC) to CBD was increased by 50% in subjects with mild hepatic impairment, 2.5-fold in subjects with moderate, and 5-fold in severe hepatic impairment.²²¹

There is debate in the literature on whether CBD potentiates or antagonises analgesic effects produced by THC. One hypothesis is that CBD is an inhibitor of CYP450 enzyme systems and may affect the pharmacokinetic profile of THC. Cannabidiol does inhibit microsomal CYP1A2, CYP2C9, CYP2C19, CYP2D6, and CYP3A4 enzymes, but seems to do so at much higher concentrations than typically achieved in plasma with clinically relevant doses.¹³⁸ In addition, in a human study of oromucosal THC and CBD administration, no major differences in THC pharmacokinetics were observed in the presence or absence of coadministered CBD.¹³⁸

In summary, cannabinoids demonstrate substantial interindividual variability in pharmacokinetics, particularly in absorption processes, whether via oral or inhaled routes. This can result in differences in drug concentrations at the desired sites of action, leading to inconsistent clinical effects. Only a minority of preclinical studies of cannabinoids and endocannabinoid system modulators in animal pain models have assessed drug pharmacokinetics. Particular attention is required in the future studies to understand the pharmacokinetic–pharmacodynamic relationships of drugs that interact with the endocannabinoid system, to improve translational success.

8. Discussion, conclusions, and future perspectives

Preclinical laboratory animal models provide an opportunity to investigate molecular and cellular changes associated with cannabinoid-based interventions. Pharmacological, electrophysiological, genetic, and optogenetic methodologies can facilitate understanding of the involvement of the endocannabinoid system in neural circuits and the opportunities to modulate the system to interfere with nociception and pain behaviour.

Key differences exist between preclinical and clinical evaluations of cannabinoids, CBMs, and endocannabinoid system modulators. Substantial evidence from the preclinical literature supports the hypothesis of cannabinoid-induced analgesia at multiple levels of analysis,²¹⁵ whereas there is less evidence of efficacy in human patients with pain and evidence is identified to be of low or very low quality.⁷⁹ Foremost amongst these differences is that the clinical and preclinical literatures generally evaluate different compounds. For example, preclinical animal studies rarely test cannabis itself, whereas most human studies use whole cannabis or cannabis extracts containing THC and CBD in various ratios or doses and a limited number of other cannabinoids (eg, THC or synthetic THC). Indeed, very few preclinical animal studies have evaluated vaporised/inhaled THC; none of the preclinical studies included in our meta-analysis assessed the effects of THC or cannabis administered in vaporised/inhaled form, and only 38 preclinical studies have administered cannabis extracts via other routes.²¹⁵ A recent preclinical brain imaging study showed that inhaled vaporised cannabis plant enriched in THC (10.3% THC; 0.05% CBD) uncoupled brain resting-state functional connectivity in the raphe nuclei in paclitaxel-treated rats, normalizing paclitaxel-induced hyperconnectivity to levels observed in vehicle-treated rats, and also produced antinociception in the cold plate test.⁷ The paucity of such studies may reflect difficulty in administering cannabis plant material to rats and mice in a manner where the dose of THC and other phytocannabinoids administered would be precise. Preclinical animal studies have performed mechanistic evaluations using individual cannabinoids and endocannabinoid system modulators (or sometimes combinations of 2 or more of these) at very well-defined doses and via routes of administration (mostly parenteral) that yield predictable pharmacokinetics.

Second, a much wider variety of cannabinoids and endocannabinoid system modulators have been tested in animal models of pain than have been assessed in human pain patients. This point is underscored by comparing our IASP Task Force clinical and preclinical systemic reviews, which reviewed 11⁷⁹ and 171²¹⁵ pharmacological interventions, respectively. Ethical, safety, and methodological standpoints require that novel/experimental drugs are administered in rodent models before human subjects and where mechanism of action can also be identified. Significant barriers to performing clinical research with cannabis and cannabinoids also remain. There are, collectively, over 150 preclinical studies assessing the efficacy of CB₂ receptor agonists, endocannabinoid reuptake inhibitors, MGL inhibitors, and FAAH inhibitors in animal models of pain, but not a single randomised controlled clinical trial of the first 3 drug classes, and only 3 randomised controlled clinical trials assessing the efficacy of FAAH inhibitors in patients with pain.^{19,123,232} This phenomenon, wherein the clinical studies lag significantly behind the preclinical studies, is of course to be expected and is certainly not unique to the cannabinoid field. However, cannabis and cannabinoid use in humans has bypassed the usual preclinical and clinical studies typical of conventional drug development because of the widespread illicit, or more recently licit, use of cannabis by the public. The disparity that exists between the variety of cannabinoids and endocannabinoid system modulators that has been tested in animal models of pain vs what has been assessed in human pain patients should be kept in mind when comparing or contrasting efficacy of specific cannabinoids in animal models vs cannabis in human patients (in addition to translational relevance of the animal model and appropriateness of the clinical indication).

Another key difference between preclinical and clinical studies is that the latter very often administer the cannabinoid or cannabis as adjunctive/add-on treatment, alongside other analgesics that the patient is using. By contrast, preclinical studies in animals almost always investigate the cannabinoid drug against a “clean” background, in the absence of any other analgesics (unless drug–drug interactions are specifically investigated). When administered adjunctively, the “window” or opportunity to see therapeutic efficacy of the cannabinoid/cannabis may well be smaller than if it were investigated against a “clean” background; if the other analgesics are already having partial effects in lowering pain, it may be more difficult for an add-on cannabinoid/cannabis to induce a further measurable reduction superimposed upon the preexisting reduction.

Our accompanying systematic review and meta-analysis provides a comprehensive summary of studies in which cannabinoids, CBMs, and endocannabinoid system modulators were assessed for antinociceptive efficacy in animal models of injury-related or pathological persistent pain.²¹⁵ The overall behavioural data effect sizes are significant, and it is clear that there is a spectrum of efficacy dependent on drug and pain model type, and also on species, strain, and other experimental factors.

The risk of internal and external validity-related biases confounding these data and exaggerating effect sizes is uncertain, but not insignificant. This could be better ascertained if future preclinical studies were reported with sufficient transparency to ascertain the methods that were used to mitigate against such biases.^{8,121,140,143,201,203} There is a need for preclinical living systematic reviews that incorporate new evidence as it becomes available,⁷¹ and closer, multidisciplinary, cross-sector collaboration to ensure that laboratory animal studies have sufficient rigor to identify promising candidate drugs and more accurately inform clinical trial design.

Several intervention strategies offer the potential for analgesia while also circumventing adverse side effects of direct-acting CB₁ agonists. These include cannabinoid CB₂ agonists, peripherally restricted CB₁ agonists, inhibitors of endocannabinoid deactivation (especially FAAH inhibitors), synergism between cannabinoids and other analgesics, mode of drug delivery (eg, local, targeted delivery), biased agonism, CB₁ PAMs, and CBD. Fatty acid amide hydrolase represents a promising target whose engagement is likely to show a reduced pattern of unwanted CB₁-mediated effects, and produce anxiolytic effects and opioid-sparing effects without being inherently reinforcing. Moreover, the recent case report of a woman with a mutation in the FAAH gene who does not experience pain or anxiety provides further support that FAAH inhibitors may not only exhibit reduced adverse side effects compared with conventional analgesic but may also exhibit additional desirable therapeutic properties (ie, reduction of stress, fear, and anxiety).^{64,160} With regards to reduction of abuse liability, CB₂ receptor agonists deserve special mention for their potential to produce antiaddiction effects.^{9,17,130,175,212,249} Cannabidiol represents another promising therapeutic agent that does not produce THC-like effects in humans or animals.^{95,127,173} The mechanism underlying therapeutic effects of CBD remains incompletely understood.^{52,95} Preclinical studies, nonetheless, suggest that CBD may attenuate opioid addiction,^{124,125} THC withdrawal,¹⁶⁸ reduce nausea,^{154,185} suppress inflammation and nociceptive behavior,^{9,17,43,195,243} and attenuate stress responding.^{14,16,33,47,72,119,136,149} Further preclinical studies of underlying mechanisms as well as clinical trials of CBD are warranted to better ascertain the therapeutic effects that may be exploited as adjunct or unitary therapy. There is also a pressing requirement to better understand the

pharmacology and therapeutic potential of the other phytocannabinoids in the cannabis plant beyond THC and CBD, and whether/how these may interact with THC and CBD in the context of analgesia. Given the substantial interindividual differences in cannabinoid pharmacokinetics among humans, further research is needed to improve the consistency of drug delivery methods, for better translation of preclinical developments to human studies. Preclinical laboratory animal models are evolving to capture aspects associated with pathological pain states observed clinically, eg, spontaneous pain, anxiety, stress, and depression comorbidities, conditions under which dynamic changes in endocannabinoid tone, enzyme activity, or receptors may be observed (for review, see Refs. 42, 76, 80, 90, 165, and 187). Consequently, future preclinical studies in laboratory animals, which evaluate novel cannabinoid interventions, reference analgesics, as well as drug development candidates that previously failed for efficacy, are required to validate treatments, understand mechanisms of action, and enhance clinical translation.

Conflict of interest statement

D. P. Finn reports an Industry-Academia research grant from Alkermes Inc and Science Foundation Ireland outside of the submitted work. He also reports research grants in the area of cannabinoids or the endocannabinoid system from Shionogi Ltd (Shionogi Science Programme), from B. Braun Ltd jointly with Science Foundation Ireland, and from the Irish Research Council, CNPq Brazil and EU INTERREG Programmes. S. Haroutounian reports grants from Disarm Therapeutics, and advisory board/consultancy fees from Vertex Pharmaceuticals, Medoc Ltd, and Rafa laboratories, outside the submitted work. A.G. Hohmann reports grants from the National Institutes of Health (National Institute on Drug Abuse, National Cancer Institute, National Institute of Neurological Diseases and Stroke), Indiana Addiction Grand Challenge, and is a coinventor on a provisional patent related to CB₂-opioid interactions. A. S.C. Rice is a Council Member of IASP and Chair of the Presidential Task Force of the IASP, undertook consultancy and advisory board work for Imperial College Consultants in the last 24 months; this has included personally remunerated work outside of the submitted work for Abide, Confo, Vertex, Pharmanovo, Lateral, Novartis, Mundipharma, Orion, Asahi Kasei, Toray & Theranexis. He was the owner of share options in Spinifex Pharmaceuticals from which personal benefit accrued between 2015 and 2019 upon the acquisition of Spinifex by Novartis. A.S.C. Rice is a named inventor on the patents—A. S.C. Rice, S. Vandevoorde, and D.M. Lambert. Methods using N-(2propenyl)hexadecanamide and related amides to relieve pain. WO2005/079771 pending, and Okuse K. et al. Methods of treating pain by inhibition of vgf activity EP13702262.0/WO2013110945. During the conduct of the study Imperial College received grants funding to support Prof Rice’s programme of research from Biotechnology and Biological Sciences Research Council (BBSRC), Medical Research Council (MRC), Wellcome Trust, Alan and Sheila Diamond Charitable Trust, British Pain Society, Royal British Legion and the European Commission (IMI2 [EQIPD]; FP7 [Neuropain] and H2020 [Dolorisk]). The remaining authors have no conflicts of interest to declare.

Acknowledgements

This work is part of the effort of the IASP Presidential Task Force on cannabis and cannabinoid analgesia. The authors would like to

thank Dr Emer Power and Darragh Mattimoe for assistance with preparation of figures, and Dr Christopher Eccleston and Dr Emma Fisher for comments on the manuscript. The International Association for the Study of Pain commissioned this work in the form of a Presidential Task Force and funded attendance for the authors at a working meeting in Washington DC, November 2019.

Article history:

Received 18 January 2021

Received in revised form 10 March 2021

Accepted 10 March 2021

Available online 15 March 2021

References

- Abbott FV, Franklin KB, Westbrook RF. The formalin test: scoring properties of the first and second phases of the pain response in rats. *PAIN* 1995;60:91–102.
- Adam JM, Clark JK, Davies K, Everett K, Fields R, Francis S, Jeremiah F, Kiyoi T, Maidment M, Morrison A, Ratcliffe P, Prosser A, Schulz J, Wishart G, Baker J, Boyce S, Campbell R, Cottney JE, Deehan M, Martin I. Low brain penetrant CB1 receptor agonists for the treatment of neuropathic pain. *Bioorg Med Chem Lett* 2012;22:2932–7.
- Adamson Barnes NS, Mitchell VA, Kazantzis NP, Vaughan CW. Actions of the dual FAAH/MAGL inhibitor JZL195 in a murine neuropathic pain model. *Br J Pharmacol* 2016;173:77–87.
- Ahmed AI, van den Elsen GA, Colbers A, van der Marck MA, Burger DM, Feuth TB, Rikkert MG, Kramers C. Safety and pharmacokinetics of oral delta-9-tetrahydrocannabinol in healthy older subjects: a randomized controlled trial. *Eur Neuropsychopharmacol* 2014;24:1475–82.
- Ahn K, Smith SE, Liimatta MB, Beidler D, Sadagopan N, Dudley DT, Young T, Wren P, Zhang Y, Swaney S, Van Becelaere K, Blankman JL, Nomura DK, Bhattachar SN, Stiff C, Nomanbhoy TK, Weerapana E, Johnson DS, Cravatt BF. Mechanistic and pharmacological characterization of PF-04457845: a highly potent and selective fatty acid amide hydrolase inhibitor that reduces inflammatory and noninflammatory pain. *J Pharmacol Exp Ther* 2011;338:114–24.
- Alexander SP, Kendall DA. The complications of promiscuity: endocannabinoid action and metabolism. *Br J Pharmacol* 2007;152:602–23.
- Alkisar I, Miller AR, Hohmann AG, Sadaka A, Cai X, Kulkarni P, Ferris CF. Inhaled cannabis suppresses chemotherapy-induced neuropathic nociception by decoupling the raphe nucleus: a functional imaging study in rats. *Biol Psychiatry Cogn Neurosci Neuroimaging* 2020. doi:10.1016/j.bpsc.2020.11.015. In Press.
- Andrews NA, Latremoliere A, Basbaum AI, Mogil JS, Porreca F, Rice AS, Woolf CJ, Currie GL, Dworkin RH, Eisenach JC, Evans S, Gewandter JS, Gover TD, Handwerker H, Huang W, Iyengar S, Jensen MP, Kennedy JD, Lee N, Levine J, Lidster K, Machin I, McDermott MP, McMahon SB, Price TJ, Ross SE, Scherrer G, Seal RP, Sena ES, Silva E, Stone L, Svensson CI, Turk DC, Whiteside G. Ensuring transparency and minimization of methodologic bias in preclinical pain research: PPRECISE considerations. *PAIN* 2016;157:901–9.
- Atwal N, Casey SL, Mitchell VA, Vaughan CW. THC and gabapentin interactions in a mouse neuropathic pain model. *Neuropharmacology* 2019;144:115–21.
- Battista N, Di Tommaso M, Bari M, Maccarrone M. The endocannabinoid system: an overview. *Front Behav Neurosci* 2012;6:9.
- Berdyshev EV. Cannabinoid receptors and the regulation of immune response. *Chem Phys Lipids* 2000;108:169–90.
- Berman P, Futoran K, Lewitus GM, Mukha D, Benami M, Shlomi T, Meiri D. A new ESI-LC/MS approach for comprehensive metabolic profiling of phytocannabinoids in Cannabis. *Sci Rep* 2018;8:14280.
- Bisogno T, Ligresti A, Di Marzo V. The endocannabinoid signalling system: biochemical aspects. *Pharmacol Biochem Behav* 2005;81:224–38.
- Bitencourt RM, Takahashi RN. Cannabidiol as a therapeutic alternative for post-traumatic stress disorder: from bench research to confirmation in human trials. *Front Neurosci* 2018;12:502.
- Blankman JL, Simon GM, Cravatt BF. A comprehensive profile of brain enzymes that hydrolyze the endocannabinoid 2-arachidonoylglycerol. *Chem Biol* 2007;14:1347–56.
- Blessing EM, Steenkamp MM, Manzanares J, Marmar CR. Cannabidiol as a potential treatment for anxiety disorders. *Neurotherapeutics* 2015;12:825–36.
- Booz GW. Cannabidiol as an emergent therapeutic strategy for lessening the impact of inflammation on oxidative stress. *Free Radic Biol Med* 2011;51:1054–61.
- Bouaboula M, Poinot-Chazel C, Bourrie B, Canat X, Calandra B, Rinaldi-Carmona M, Le Fur G, Casellas P. Activation of mitogen-activated protein kinases by stimulation of the central cannabinoid receptor CB1. *Biochem J* 1995;312:637–41.
- Bradford D, Stirling A, Ernault E, Liosatos M, Tracy K, Moseley J, Blahunka P, Smith MD. The MOBILE study—A phase IIa enriched enrollment randomized withdrawal trial to assess the analgesic efficacy and safety of ASP8477, a fatty acid amide hydrolase inhibitor, in patients with peripheral neuropathic pain. *Pain Med* 2017;18:2388–400.
- Brenneman DE, Petkanas D, Kinney WA. Pharmacological comparisons between cannabidiol and KLS-13019. *J Mol Neurosci* 2018;66:121–34.
- Bridges D, Rice AS, Egertova M, Elphick MR, Winter J, Michael GJ. Localisation of cannabinoid receptor 1 in rat dorsal root ganglion using *in situ* hybridisation and immunohistochemistry. *Neuroscience* 2003;119:803–12.
- Britch SC, Babalonis S, Walsh SL. Cannabidiol: pharmacology and therapeutic targets. *Psychopharmacology (Berl)* 2020.
- Brown AJ. Novel cannabinoid receptors. *Br J Pharmacol* 2007;152:567–75.
- Brunet B, Doucet C, Venisse N, Hauet T, Hébrard W, Papet Y, Mauco G, Mura P. Validation of Large White Pig as an animal model for the study of cannabinoids metabolism: application to the study of THC distribution in tissues. *Forensic Sci Int* 2006;161:169–74.
- Bruni N, Della Pepa C, Oliaro-Bosso S, Pessione E, Gastaldi D, Dosio F. Cannabinoid delivery systems for pain and inflammation treatment. *Molecules* 2018;23:2478.
- Bujalska-Zadrozny M, de Corde A, Pawlik K. Influence of nitric oxide synthase or cyclooxygenase inhibitors on cannabinoids activity in streptozotocin-induced neuropathy. *Pharmacol Rep* 2015;67:209–16.
- Butler RK, Finn DP. Stress-induced analgesia. *Prog Neurobiol* 2009;88:184–202.
- Butler RK, Rea K, Lang Y, Gavin AM, Finn DP. Endocannabinoid-mediated enhancement of fear-conditioned analgesia in rats: opioid dependency and molecular correlates. *PAIN* 2008;140:491–500.
- Cabanero D, Ramirez-Lopez A, Drews E, Schmöle A, Otte DM, Wawrzczak-Bargiela A, Huerga Encabo R, Kummer S, Ferrer-Montiel A, Przewlocki R, Zimmer A, Maldonado H. Protective role of neuronal and lymphoid cannabinoid CB2 receptors in neuropathic pain. *Elife*;9:2020.
- Cadas H, di Tomaso E, Piomelli D. Occurrence and biosynthesis of endogenous cannabinoid precursor, N-arachidonoyl phosphatidylethanolamine, in rat brain. *J Neurosci* 1997;17:1226–42.
- Cadas H, Gaillet S, Beltramo M, Venance L, Piomelli D. Biosynthesis of an endogenous cannabinoid precursor in neurons and its control by calcium and cAMP. *J Neurosci* 1996;16:3934–42.
- Calignano A, La Rana G, Giuffrida A, Piomelli D. Control of pain initiation by endogenous cannabinoids. *Nature* 1998;394:277–81.
- Campos AC, Ferreira FR, Guimaraes FS. Cannabidiol blocks long-lasting behavioral consequences of predator threat stress: possible involvement of 5HT1A receptors. *J Psychiatry Res* 2012;46:1501–10.
- Casey SL, Atwal N, Vaughan CW. Cannabis constituent synergy in a mouse neuropathic pain model. *PAIN* 2017;158:2452–60.
- Chen JP, Paredes W, Li J, Smith D, Lowinson J, Gardner EL. Delta 9-tetrahydrocannabinol produces naloxone-blockable enhancement of presynaptic basal dopamine efflux in nucleus accumbens of conscious, freely-moving rats as measured by intracerebral microdialysis. *Psychopharmacology (Berl)* 1990;102:156–62.
- Cherniakov I, Izgelov D, Barasch D, Davidson E, Domb AJ, Hoffman A. Piperine-pro-nanoliposomes as a novel oral delivery system of cannabinoids: pharmacokinetic evaluation in healthy volunteers in comparison to buccal spray administration. *J Control Release* 2017;266:1–7.
- Cherniakov I, Izgelov D, Domb AJ, Hoffman A. The effect of Pro NanoLiposomes (PNL) formulation containing natural absorption enhancers on the oral bioavailability of delta-9-tetrahydrocannabinol (THC) and cannabidiol (CBD) in a rat model. *Eur J Pharm Sci* 2017;109:21–30.
- Clapper JR, Moreno-Sanz G, Russo R, Guijarro A, Vacondio F, Duranti A, Tontini A, Sanchini S, Sciolino NR, Spradley JM, Hohmann AG, Calignano A, Mor M, Tarzia G, Piomelli D. Anandamide suppresses pain

- initiation through a peripheral endocannabinoid mechanism. *Nat Neurosci* 2010;13:1265–70.
- [39] Clayton JA, Collins FS. Policy: NIH to balance sex in cell and animal studies. *Nature* 2014;509:282–3.
- [40] Conner SN, Bedell V, Lipsey K, Macones GA, Cahill AG, Tuuli MG. Maternal marijuana use and adverse neonatal outcomes: a systematic review and meta-analysis. *Obstet Gynecol* 2016;128:713–23.
- [41] Conner SN, Carter EB, Tuuli MG, Macones GA, Cahill AG. Maternal marijuana use and neonatal morbidity. *Am J Obstet Gynecol* 2015;213:422 e421–424.
- [42] Corcoran L, Roche M, Finn DP. The role of the brain's endocannabinoid system in pain and its modulation by stress. *Int Rev Neurobiol* 2015;125:203–55.
- [43] Costa B, Trovato AE, Comelli F, Giagnoni G, Colleoni M. The non-psychoactive cannabis constituent cannabidiol is an orally effective therapeutic agent in rat chronic inflammatory and neuropathic pain. *Eur J Pharmacol* 2007;556:75–83.
- [44] Cox ML, Haller VL, Welch SP. The antinociceptive effect of Delta9-tetrahydrocannabinol in the arthritic rat involves the CB(2) cannabinoid receptor. *Eur J Pharmacol* 2007;570:50–6.
- [45] Cox ML, Haller VL, Welch SP. Synergy between delta9-tetrahydrocannabinol and morphine in the arthritic rat. *Eur J Pharmacol* 2007;567:125–30.
- [46] Cravatt BF, Giang DK, Mayfield SP, Boger DL, Lerner RA, Gilula NB. Molecular characterization of an enzyme that degrades neuromodulatory fatty-acid amides. *Nature* 1996;384:83–7.
- [47] Crippa JA, Guimaraes FS, Campos AC, Zuardi AW. Translational investigation of the therapeutic potential of cannabidiol (CBD): toward a new age. *Front Immunol* 2009;9:2018.
- [48] Crossley NA, Sena E, Goehler J, Horn J, van der Worp B, Bath PM, Macleod M, Dirnagl U. Empirical evidence of bias in the design of experimental stroke studies: a metaepidemiologic approach. *Stroke* 2008;39:929–34.
- [49] Crowe MS, Leishman E, Banks ML, Gujjar R, Mahadevan A, Bradshaw HB, Kinsey SG. Combined inhibition of monoacylglycerol lipase and cyclooxygenases synergistically reduces neuropathic pain in mice. *Br J Pharmacol* 2015;172:1700–12.
- [50] Curry ZA, Wilkerson JL, Bagdas D, Kyte SL, Patel N, Donvito G, Mustafa MA, Poklis JL, Niphakis MJ, Hsu KL, Cravatt BF, Gewirtz DA, Damaj MI, Lichtman AH. Monoacylglycerol lipase inhibitors reverse paclitaxel-induced nociceptive behavior and proinflammatory markers in a mouse model of chemotherapy-induced neuropathy. *J Pharmacol Exp Ther* 2018;366:169–83.
- [51] D'Souza DC, Cortes-Briones J, Creatura G, Bluez G, Thurnauer H, Deaso E, Bielen K, Surti T, Radhakrishnan R, Gupta A, Gupta S, Cahill J, Sherif MA, Makriyannis A, Morgan PT, Ranganathan M, Skosnik PD. Efficacy and safety of a fatty acid amide hydrolase inhibitor (PF-04457845) in the treatment of cannabis withdrawal and dependence in men: a double-blind, placebo-controlled, parallel group, phase 2a single-site randomised controlled trial. *Lancet Psychiatry* 2019;6:35–45.
- [52] De Gregorio D, McLaughlin RJ, Posa L, Ochoa-Sanchez R, Enns J, Lopez-Canul M, Aboud M, Maione S, Comai S, Gobbi G. Cannabidiol modulates serotonergic transmission and reverses both allodynia and anxiety-like behavior in a model of neuropathic pain. *PAIN* 2019;160:136–50.
- [53] Deciga-Campos M, Ortiz-Andrade R. Enhancement of antihyperalgesia by the coadministration of N-palmitoylethanolamide and acetaminophen in diabetic rats. *Drug Dev Res* 2015;76:228–34.
- [54] Deiana S, Watanabe A, Yamasaki Y, Amada N, Arthur M, Fleming S, Woodcock H, Dorward P, Pigliacampo B, Close S, Platt B, Riedel G. Plasma and brain pharmacokinetic profile of cannabidiol (CBD), cannabidivarin (CBDV), Delta(9)-tetrahydrocannabivarin (THCV) and cannabigerol (CBG) in rats and mice following oral and intraperitoneal administration and CBD action on obsessive-compulsive behaviour. *Psychopharmacology (Berl)* 2012;219:859–73.
- [55] Demuth DG, Molleman A. Cannabinoid signalling. *Life Sci* 2006;78:549–63.
- [56] Deng L, Cornett BL, Mackie K, Hohmann AG. CB1 knockout mice unveil sustained CB2-mediated antialloodynic effects of the mixed CB1/CB2 agonist CP55,940 in a mouse model of paclitaxel-induced neuropathic pain. *Mol Pharmacol* 2015;88:64–74.
- [57] Devane WA, Dysarz FA III, Johnson MR, Melvin LS, Howlett AC. Determination and characterization of a cannabinoid receptor in rat brain. *Mol Pharmacol* 1988;34:605–13.
- [58] Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, Gibson D, Mandelbaum A, Etinger A, Mechoulam R. Isolation and structure of a brain constituent that binds to the cannabinoid receptor [see comments]. *Science* 1992;258:1946–9.
- [59] Di Marzo V. Endocannabinoids: synthesis and degradation. *Rev Physiol Biochem Pharmacol* 2008;160:1–24.
- [60] Di Marzo V. Targeting the endocannabinoid system: to enhance or reduce? *Nat Rev Drug Discov* 2008;7:438–55.
- [61] Di Marzo V, Breivogel CS, Tao Q, Bridgen DT, Razdan RK, Zimmer AM, Zimmer A, Martin BR. Levels, metabolism, and pharmacological activity of anandamide in CB(1) cannabinoid receptor knockout mice: evidence for non-CB(1), non-CB(2) receptor-mediated actions of anandamide in mouse brain. *J Neurochem* 2000;75:2434–44.
- [62] Di Marzo V, Fontana A, Cadas H, Schinelli S, Cimino G, Schwartz JC, Piomelli D. Formation and inactivation of endogenous cannabinoid anandamide in central neurons. *Nature* 1994;372:686–91.
- [63] Di Marzo V, Stella N, Zimmer A. Endocannabinoid signalling and the deteriorating brain. *Nat Rev Neurosci* 2015;16:30–42.
- [64] Dincheva I, Drysdale AT, Hartley CA, Johnson DC, Jing D, King EC, Ra S, Gray JM, Yang R, DeGruccio AM, Huang C, Cravatt BF, Glatt CE, Hill MN, Casey BJ, Lee FS. FAAH genetic variation enhances frontoamygdala function in mouse and human. *Nat Commun* 2015;6:6395.
- [65] Dinh TP, Carpenter D, Leslie FM, Freund TF, Katona I, Sensi SL, Kathuria S, Piomelli D. Brain monoglyceride lipase participating in endocannabinoid inactivation. *Proc Natl Acad Sci U S A* 2002;99:10819–24.
- [66] Donvito G, Nass SR, Wilkerson JL, Curry ZA, Schurman LD, Kinsey SG, Lichtman AH. The endogenous cannabinoid system: a budding source of targets for treating inflammatory and neuropathic pain. *Neuropsychopharmacology* 2018;43:52–79.
- [67] Donvito G, Wilkerson JL, Damaj MI, Lichtman AH. Palmitoylethanolamide reverses paclitaxel-induced allodynia in mice. *J Pharmacol Exp Ther* 2016;359:310–18.
- [68] Dos Santos RG, Hallak JEC, Crippa JAS. Neuropharmacological effects of the main phytocannabinoids: a narrative review. *Adv Exp Med Biol* 2021;1264:29–45.
- [69] Dubuisson D, Dennis SG. The formalin test: a quantitative study of the analgesic effects of morphine, meperidine, and brain stem stimulation in rats and cats. *PAIN* 1977;4:161–74.
- [70] Eisenberg E, Ogintz M, Almog S. The pharmacokinetics, efficacy, safety, and ease of use of a novel portable metered-dose cannabis inhaler in patients with chronic neuropathic pain: a phase 1a study. *J Pain Palliat Care Pharmacother* 2014;28:216–25.
- [71] Elliott JH, Synnot A, Turner T, Simmonds M, Akl EA, McDonald S, Salanti G, Meerpohl J, MacLehose H, Hilton J, Tovey D, Shemilt I, Thomas J. Living Systematic Review N. Living systematic review: 1. Introduction—the why, what, when, and how. *J Clin Epidemiol* 2017;91:23–30.
- [72] Elms L, Shannon S, Hughes S, Lewis N. Cannabidiol in the treatment of post-traumatic stress disorder: a case series. *J Altern Complement Med* 2019;25:392–7.
- [73] Endo T, Takeuchi T, Maehara S. Pharmacological characterization of a novel, potent, selective, and orally active fatty acid amide hydrolase inhibitor, PKM-833 [(R)-N-(pyridazin-3-yl)-4-(7-(trifluoromethyl)chroman-4-yl)piperazine-1-carboxamide] in rats: potential for the treatment of inflammatory pain. *Pharmacol Res Perspect* 2020;8:e00569.
- [74] Farquhar-Smith WP, Egertova M, Bradbury EJ, McMahon SB, Rice AS, Elphick MR. Cannabinoid CB(1) receptor expression in rat spinal cord. *Mol Cell Neurosci* 2000;15:510–21.
- [75] Fine PG, Rosenfeld MJ. The endocannabinoid system, cannabinoids, and pain. *Rambam Maimonides Med J* 2013;4:e0022.
- [76] Finn DP. Endocannabinoid-mediated modulation of stress responses: physiological and pathophysiological significance. *Immunobiology* 2010;215:629–46.
- [77] Finn DP, Beckett SR, Richardson D, Kendall DA, Marsden CA, Chapman V. Evidence for differential modulation of conditioned aversion and fear-conditioned analgesia by CB1 receptors. *Eur J Neurosci* 2004;20:848–52.
- [78] Finn DP, Beckett SR, Roe CH, Madjid A, Fone KC, Kendall DA, Marsden CA, Chapman V. Effects of coadministration of cannabinoids and morphine on nociceptive behaviour, brain monoamines and HPA axis activity in a rat model of persistent pain. *Eur J Neurosci* 2004;19:678–86.
- [79] Fisher E, Moore RA, Fogarty AE, Finn DP, Finnerup NB, Gilron I, Haroutounian S, Krane E, Rice ASC, Rowbotham M, Wallace M, Eccleston C. Cannabinoids, cannabis, and cannabis-based medicine for pain management: a systematic review of randomised controlled trials. *PAIN* 2020. doi: 10.1097/j.pain.0000000000001929. [Epub ahead of print].
- [80] Fitzgibbon M, Finn DP, Roche M. High times for painful blues: the endocannabinoid system in pain-depression comorbidity. *Int J Neuropsychopharmacol* 2015;19:pyv095.

- [81] Fox A, Kessingland A, Gentry C, McNair K, Patel S, Urban L, James I. The role of central and peripheral Cannabinoid1 receptors in the antihyperalgesic activity of cannabinoids in a model of neuropathic pain. *PAIN* 2001;92:91–100.
- [82] Franco V, Perucca E. Pharmacological and therapeutic properties of cannabidiol for epilepsy. *Drugs* 2019;79:1435–54.
- [83] Gaoni Y, Mechoulam R. Isolation, structure and partial synthesis of an active constituent of hashish. *J Am Chem Soc* 1964;86:1646–7.
- [84] Genaro K, Fabris D, Arantes ALF, Zuardi AW, Crippa JAS, Prado WA. Cannabidiol is a potential therapeutic for the affective-motivational dimension of incision pain in rats. *Front Pharmacol* 2017;8:391.
- [85] Gershkovich P, Qadri B, Yacovan A, Amselem S, Hoffman A. Different impacts of intestinal lymphatic transport on the oral bioavailability of structurally similar synthetic lipophilic cannabinoids: dexamnabinol and PRS-211,220. *Eur J Pharm Sci* 2007;31:298–305.
- [86] Giang DK, Cravatt BF. Molecular characterization of human and mouse fatty acid amide hydrolases. *Proc Natl Acad Sci U S A* 1997;94:2238–42.
- [87] Ginsburg BC, Hruba L, Zaki A, Javors MA, McMahon LR. Blood levels do not predict behavioral or physiological effects of Δ^9 -tetrahydrocannabinol in rhesus monkeys with different patterns of exposure. *Drug Alcohol Depend* 2014;139:1–8.
- [88] Glass M, Dragunov M, Faull RLM. Cannabinoid receptors in the human brain: a detailed anatomical and quantitative autoradiographic study in the fetal, neonatal and adult human brain. *Neuroscience* 1997;77:299–318.
- [89] Goparaju SK, Ueda N, Yamaguchi H, Yamamoto S. Anandamide amidohydrolase reacting with 2-arachidonoylglycerol, another cannabinoid receptor ligand. *FEBS Lett* 1998;422:69–73.
- [90] Gorzalka BB, Hill MN. Putative role of endocannabinoid signaling in the etiology of depression and actions of antidepressants. *Prog Neuropsychopharmacol Biol Psychiatry* 2011;35:1575–85.
- [91] Grant KS, Petroff R, Isoherranen N, Stella N, Burbacher TM. Cannabis use during pregnancy: pharmacokinetics and effects on child development. *Pharmacol Ther* 2018;182:133–51.
- [92] Gregg LC, Jung KM, Spradley JM, Nyilas R, Suplita RL II, Zimmer A, Watanabe M, Mackie K, Katona I, Piomelli D, Hohmann AG. Activation of type 5 metabotropic glutamate receptors and diacylglycerol lipase- α initiates 2-arachidonoylglycerol formation and endocannabinoid-mediated analgesia. *J Neurosci* 2012;32:9457–68.
- [93] Grenald SA, Young MA, Wang Y, Ossipov MH, Ibrahim MM, Largent-Milnes TM, Vanderah TW. Synergistic attenuation of chronic pain using mu opioid and cannabinoid receptor 2 agonists. *Neuropharmacology* 2017;116:59–70.
- [94] Grotenhermen F, Muller-Vahl K. The therapeutic potential of cannabis and cannabinoids. *Dtsch Arztebl Int* 2012;109:495–501.
- [95] Grotenhermen F, Russo E, Zuardi AW. Even High Doses of Oral Cannabidiol Do Not Cause THC-Like Effects in Humans: comment on Merrick et al. *Cannabis Cannabinoid Res* 2016;1:102–12.
- [96] Guindon J, Beaulieu P. The role of the endogenous cannabinoid system in peripheral analgesia. *Curr Mol Pharmacol* 2009;2:134–9.
- [97] Guindon J, De Lean A, Beaulieu P. Local interactions between anandamide, an endocannabinoid, and ibuprofen, a nonsteroidal anti-inflammatory drug, in acute and inflammatory pain. *PAIN* 2006;121:85–93.
- [98] Guindon J, Hohmann AG. Cannabinoid CB2 receptors: a therapeutic target for the treatment of inflammatory and neuropathic pain. *Br J Pharmacol* 2008;153:319–34.
- [99] Guindon J, Hohmann AG. The endocannabinoid system and pain. *CNS Neurol Disord Drug Targets* 2009;8:403–21.
- [100] Guindon J, Lai Y, Takacs SM, Bradshaw HB, Hohmann AG. Alterations in endocannabinoid tone following chemotherapy-induced peripheral neuropathy: effects of endocannabinoid deactivation inhibitors targeting fatty-acid amide hydrolase and monoacylglycerol lipase in comparison to reference analgesics following cisplatin treatment. *Pharmacol Res* 2013;67:94–109.
- [101] Guindon J, LoVerme J, De Lean A, Piomelli D, Beaulieu P. Synergistic antinociceptive effects of anandamide, an endocannabinoid, and nonsteroidal anti-inflammatory drugs in peripheral tissue: a role for endogenous fatty-acid ethanolamides? *Eur J Pharmacol* 2006;550:68–77.
- [102] Gutierrez T, Crystal JD, Zvonok AM, Makriyannis A, Hohmann AG. Self-medication of a cannabinoid CB2 agonist in an animal model of neuropathic pain. *PAIN* 2011;152:1976–87.
- [103] Hanus LO, Meyer SM, Munoz E, Tagliatalata-Scafati O, Appendino G. Phytocannabinoids: a unified critical inventory. *Nat Prod Rep* 2016;33:1357–92.
- [104] Herkenham M, Lynn AB, Johnson MR, Melvin LS, de Costa BR, Rice KC. Characterization and localization of cannabinoid receptors in rat brain: a quantitative in vitro autoradiographic study. *J Neurosci* 1991;11:563–83.
- [105] Hirst JA, Howick J, Aronson JK, Roberts N, Perera R, Koshiaris C, Heneghan C. The need for randomization in animal trials: an overview of systematic reviews. *PLoS One* 2014;9:e98856.
- [106] Hložek T, Uttl L, Kadeřábek L, Balíková M, Lhotková E, Horsley RR, Nováková P, Šichová K, Štefková K, Tyš F, Kuchař M, Páleníček T. Pharmacokinetic and behavioural profile of THC, CBD, and THC+CBD combination after pulmonary, oral, and subcutaneous administration in rats and confirmation of conversion in vivo of CBD to THC. *Eur Neuropsychopharmacol* 2017;27:1223–37.
- [107] Hohmann AG. Spinal and peripheral mechanisms of cannabinoid antinociception: behavioral, neurophysiological and neuroanatomical perspectives. *Chem Phys Lipids* 2002;121:173–90.
- [108] Hohmann AG, Briley EM, Herkenham M. Pre- and postsynaptic distribution of cannabinoid and mu opioid receptors in rat spinal cord. *Brain Res* 1999;822:17–25.
- [109] Hohmann AG, Herkenham M. Regulation of cannabinoid and mu opioid receptors in rat lumbar spinal cord following neonatal capsaicin treatment. *Neurosci Lett* 1998;252:13–16.
- [110] Hohmann AG, Herkenham M. Cannabinoid receptors undergo axonal flow in sensory nerves. *Neuroscience* 1999;92:1171–5.
- [111] Hohmann AG, Herkenham M. Localization of central cannabinoid CB1 receptor messenger RNA in neuronal subpopulations of rat dorsal root ganglia: a double-label in situ hybridization study. *Neuroscience* 1999;90:923–31.
- [112] Hohmann AG, Martin WJ, Tsou K, Walker JM. Inhibition of noxious stimulus-evoked activity of spinal cord dorsal horn neurons by the cannabinoid WIN 55,212-2. *Life Sci* 1995;56:2111–18.
- [113] Hohmann AG, Suplita RL, Bolton NM, Neely MH, Fegley D, Mangieri R, Krey JF, Walker JM, Holmes PV, Crystal JD, Duranti A, Tontini A, Mor M, Tarzia G, Piomelli D. An endocannabinoid mechanism for stress-induced analgesia. *Nature* 2005;435:1108–12.
- [114] Hohmann AG, Tsou K, Walker JM. Cannabinoid modulation of wide dynamic range neurons in the lumbar dorsal horn of the rat by spinally administered WIN55,212-2. *Neurosci Lett* 1998;257:119–22.
- [115] Hohmann AG, Tsou K, Walker JM. Intrathecal cannabinoid administration suppresses noxious stimulus-evoked Fos protein-like immunoreactivity in rat spinal cord: comparison with morphine. *Acta Pharmacol Sin* 1999;20:1132–6.
- [116] Howlett AC. Inhibition of neuroblastoma adenylate cyclase by cannabinoid and nantradol compounds. *Life Sci* 1984;35:1803–10.
- [117] Howlett AC, Fleming RM. Cannabinoid inhibition of adenylate cyclase. Pharmacology of the response in neuroblastoma cell membranes. *Mol Pharmacol* 1984;26:532–8.
- [118] Howlett AC, Mukhopadhyay S, Shim JY, Welsh WJ. Signal transduction of eicosanoid CB1 receptor ligands. *Life Sci* 1999;65:617–25.
- [119] Hsiao YT, Yi PL, Li CL, Chang FC. Effect of cannabidiol on sleep disruption induced by the repeated combination tests consisting of open field and elevated plus-maze in rats. *Neuropharmacology* 2012;62:373–84.
- [120] Hu B, Doods H, Treede RD, Ceci A. Depression-like behaviour in rats with mononeuropathy is reduced by the CB2-selective agonist GW405833. *PAIN* 2009;143:206–12.
- [121] Huang W, Percie du Sert N, Vollert J, Rice ASC. General principles of preclinical study design. *Handb Exp Pharmacol* 2020;257:55–69.
- [122] Huestis M. Pharmacokinetics of THC in inhaled and oral preparations. In: Nahas GG, Sutin KM, Harvey DJ, Agurell S, editors. *Marihuana and Medicine*. Totowa, NJ: Humana Press, 1999, p. 105–16.
- [123] Huggins JP, Smart TS, Langman S, Taylor L, Young T. An efficient randomised, placebo-controlled clinical trial with the irreversible fatty acid amide hydrolase-1 inhibitor PF-04457845, which modulates endocannabinoids but fails to induce effective analgesia in patients with pain due to osteoarthritis of the knee. *PAIN* 2012;153:1837–46.
- [124] Hurd YL, Spriggs S, Alishayev J, Winkel G, Gurgov K, Kudrich C, Oprescu AM, Salsitz E. Cannabidiol for the reduction of cue-induced craving and anxiety in drug-abstinent individuals with heroin use disorder: a double-blind randomized placebo-controlled trial. *Am J Psychiatry* 2019;176:911–22.
- [125] Hurd YL, Yoon M, Manini AF, Hernandez S, Olmedo R, Ostman M, Jutras-Aswad D. Early phase in the development of cannabidiol as a treatment for addiction: opioid relapse takes initial center stage. *Neurotherapeutics* 2015;12:807–15.
- [126] Ibrahim MM, Deng H, Zvonok A, Cockayne DA, Kwan J, Mata HP, Vanderah TW, Lai J, Porreca F, Makriyannis A, Malan TP Jr. Activation of CB2 cannabinoid receptors by AM1241 inhibits experimental

- neuropathic pain: pain inhibition by receptors not present in the CNS. *Proc Natl Acad Sci U S A* 2003;100:10529–33.
- [127] Iffland K, Grotenhermen F. An update on safety and side effects of cannabidiol: a review of clinical data and relevant animal studies. *Cannabis Cannabinoid Res* 2017;2:139–54.
- [128] Ignatowska-Jankowska B, Wilkerson JL, Mustafa M, Abdullah R, Niphakis M, Wiley JB, Cravatt BF, Lichtman AH. Selective monoacylglycerol lipase inhibitors: antinociceptive versus cannabimimetic effects in mice. *J Pharmacol Exp Ther* 2015;353:424–32.
- [129] Ignatowska-Jankowska BM, Baillie GL, Kinsey S, Crowe M, Ghosh S, Owens RA, Damaj IM, Poklis J, Wiley JL, Zanda M, Zanato C, Greig IR, Lichtman AH, Ross RA. A cannabinoid CB1 receptor-positive allosteric modulator reduces neuropathic pain in the mouse with no psychoactive effects. *Neuropsychopharmacology* 2015;40:2948–59.
- [130] Iyer V, Slivicki RA, Thomaz AC, Crystal JD, Mackie K, Hohmann AG. The cannabinoid CB2 receptor agonist LY2828360 synergizes with morphine to suppress neuropathic nociception and attenuates morphine reward and physical dependence. *Eur J Pharmacol* 2020;886:173544.
- [131] Izgelov D, Regev A, Domb AJ, Hoffman A. Using the absorption cocktail approach to assess differential absorption kinetics of cannabidiol administered in lipid-based vehicles in rats. *Mol Pharm* 2020;17:1979–86.
- [132] Jiang HX, Ke BW, Liu J, Ma G, Hai KR, Gong DY, Yang Z, Zhou C. Inhibition of fatty acid amide hydrolase improves depressive-like behaviors independent of its peripheral antinociceptive effects in a rat model of neuropathic pain. *Anesth Analg* 2019;129:587–97.
- [133] Jonsson KO, Vandevoorde S, Lambert DM, Tiger G, Fowler CJ. Effects of homologues and analogues of palmitoylethanolamide upon the inactivation of the endocannabinoid anandamide. *Br J Pharmacol* 2001;133:1263–75.
- [134] Jordan CJ, Xi ZX. Progress in brain cannabinoid CB2 receptor research: from genes to behavior. *Neurosci Biobehav Rev* 2019;98:208–20.
- [135] Jung KM, Astarita G, Zhu C, Wallace M, Mackie K, Piomelli D. A key role for diacylglycerol lipase- α in metabotropic glutamate receptor-dependent endocannabinoid mobilization. *Mol Pharmacol* 2007;72:612–21.
- [136] Jurkus R, Day HL, Guimaraes FS, Lee JL, Bertoglio LJ, Stevenson CW. Cannabidiol regulation of learned fear: implications for treating anxiety-related disorders. *Front Pharmacol* 2016;7:454.
- [137] Karler R, Sangdee P, Turkanis SA, Borys HK. The pharmacokinetic fate of cannabidiol and its relationship to barbiturate sleep time. *Biochem Pharmacol* 1979;28:777–84.
- [138] Karschner EL, Darwin WD, Goodwin RS, Wright S, Huestis MA. Plasma cannabinoid pharmacokinetics following controlled oral delta9-tetrahydrocannabinol and oromucosal cannabis extract administration. *Clin Chem* 2011;57:66–75.
- [139] Khurana L, Mackie K, Piomelli D, Kendall DA. Modulation of CB1 cannabinoid receptor by allosteric ligands: pharmacology and therapeutic opportunities. *Neuropharmacology* 2017;124:3–12.
- [140] Kilkeny C, Browne W, Cuthill IC, Emerson M, Altman DG, Group NCRGW. Animal research: reporting in vivo experiments: the ARRIVE guidelines. *Br J Pharmacol* 2010;160:1577–9.
- [141] King KM, Myers AM, Soroka-Monzo AJ, Tuma RF, Tallarida RJ, Walker EA, Ward SJ. Single and combined effects of Delta(9)-tetrahydrocannabinol and cannabidiol in a mouse model of chemotherapy-induced neuropathic pain. *Br J Pharmacol* 2017;174:2832–41.
- [142] Kinsey SG, Wise LE, Ramesh D, Abdullah R, Selley DE, Cravatt BF, Lichtman AH. Repeated low-dose administration of the monoacylglycerol lipase inhibitor JZL184 retains cannabinoid receptor type 1-mediated antinociceptive and gastroprotective effects. *J Pharmacol Exp Ther* 2013;345:492–501.
- [143] Knopp KL, Stenfors C, Baastrup C, Bannon AW, Calvo M, Caspani O, Currie G, Finnerup NB, Huang W, Kennedy JD, Lefevre I, Machin I, Macleod M, Rees H, Rice ASC, Rutten K, Segerdahl M, Serra J, Wodarski R, Berge OG, Treede RD. Experimental design and reporting standards for improving the internal validity of pre-clinical studies in the field of pain: consensus of the IMI-Europain consortium. *Scand J Pain* 2015;7:58–70.
- [144] Kreitzer AC, Regehr WG. Retrograde inhibition of presynaptic calcium influx by endogenous cannabinoids at excitatory synapses onto Purkinje cells. *Neuron* 2001;29:717–27.
- [145] Kwilasz AJ, Abdullah RA, Poklis JL, Lichtman AH, Negus SS. Effects of the fatty acid amide hydrolase inhibitor URB597 on pain-stimulated and pain-depressed behavior in rats. *Behav Pharmacol* 2014;25:119–29.
- [146] Kwilasz AJ, Negus SS. Dissociable effects of the cannabinoid receptor agonists Delta9-tetrahydrocannabinol and CP55940 on pain-stimulated versus pain-depressed behavior in rats. *J Pharmacol Exp Ther* 2012;343:389–400.
- [147] La Porta C, Bura SA, Llorente-Onaindia J, Pastor A, Navarrete F, Garcia-Gutierrez MS, De la Torre R, Manzanares J, Monfort J, Maldonado R. Role of the endocannabinoid system in the emotional manifestations of osteoarthritis pain. *PAIN* 2015;156:2001–12.
- [148] Ledent C, Valverde O, Cossu G, Petitot F, Aubert JF, Beslot F, Bohme GA, Imperato A, Pedrazzini T, Roques BP, Vassart G, Fratta W, Parmentier M. Unresponsiveness to cannabinoids and reduced addictive effects of opiates in CB1 receptor knockout mice. *Science* 1999;283:401–4.
- [149] Lee JLC, Bertoglio LJ, Guimaraes FS, Stevenson CW. Cannabidiol regulation of emotion and emotional memory processing: relevance for treating anxiety-related and substance abuse disorders. *Br J Pharmacol* 2017;174:3242–56.
- [150] Leitl MD, Negus SS. Pharmacological modulation of neuropathic pain-related depression of behavior: effects of morphine, ketoprofen, bupropion and [INCREMENT]9-tetrahydrocannabinol on formalin-induced depression of intracranial self-stimulation in rats. *Behav Pharmacol* 2016;27:364–76.
- [151] Lepore M, Vorel SR, Lowinson J, Gardner EL. Conditioned place preference induced by delta 9-tetrahydrocannabinol: comparison with cocaine, morphine, and food reward. *Life Sci* 1995;56:2073–80.
- [152] Lever IJ, Pheby TM, Rice AS. Continuous infusion of the cannabinoid WIN 55,212-2 to the site of a peripheral nerve injury reduces mechanical and cold hypersensitivity. *Br J Pharmacol* 2007;151:292–302.
- [153] Li AL, Lin X, Dhopeswarkar AS, Thomaz AC, Carey LM, Liu Y, Nikas SP, Makriyannis A, Mackie K, Hohmann AG. Cannabinoid CB2 agonist AM1710 differentially suppresses distinct pathological pain states and attenuates morphine tolerance and withdrawal. *Mol Pharmacol* 2019;95:155–68.
- [154] Limebeer CL, Rock EM, Sharkey KA, Parker LA. Nausea-induced 5-HT release in the interoceptive insular cortex and regulation by monoacylglycerol lipase (MAGL) inhibition and cannabidiol. *eNeuro* 2018;5:ENEURO.0256-18.2018.
- [155] Lin X, Dhopeswarkar AS, Huijbreghse M, Mackie K, Hohmann AG. Slowly signaling G protein-biased CB2 cannabinoid receptor agonist LY2828360 suppresses neuropathic pain with sustained efficacy and attenuates morphine tolerance and dependence. *Mol Pharmacol* 2018;93:49–62.
- [156] Little PJ, Compton DR, Johnson MR, Melvin LS, Martin BR. Pharmacology and stereoselectivity of structurally novel cannabinoids in mice. *J Pharmacol Exp Ther* 1988;247:1046–51.
- [157] Luongo L, Maione S, Di Marzo V. Endocannabinoids and neuropathic pain: focus on neuron-glia and endocannabinoid-neurotrophin interactions. *Eur J Neurosci* 2014;39:401–8.
- [158] Martin WJ, Hohmann AG, Walker JM. Suppression of noxious stimulus-evoked activity in the ventral posterolateral nucleus of the thalamus by a cannabinoid agonist: correlation between electrophysiological and antinociceptive effects. *J Neurosci* 1996;16:6601–11.
- [159] Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI. Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* 1990;346:561–4.
- [160] Mayo LM, Asratian A, Linde J, Morena M, Haataja R, Hammar V, Augier G, Hill MN, Heilig M. Elevated anandamide, enhanced recall of fear extinction, and attenuated stress responses following inhibition of fatty acid amide hydrolase: a randomized, controlled experimental medicine trial. *Biol Psychiatry* 2019.
- [161] Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, Gopher A, Almog S, Martin BR, Compton DR. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem Pharmacol* 1995;50:83–90.
- [162] Mechoulam R, Gaoni Y. Isolation, structure and partial synthesis of an active constituent of hashish. *J Am Chem Soc* 1967;86:1646–7.
- [163] Miederer I, Buchholz HG, Kronfeld A, Maus S, Weyer-Elberich V, Mildenerberger P, Lutz B, Schreckenberger M. Pharmacokinetics of the cannabinoid receptor ligand [¹⁸F]JMK-9470 in the rat brain—Evaluation of models using microPET. *Med Phys* 45:725–34, 2018.
- [164] Mogil JS. Sex differences in pain and pain inhibition: multiple explanations of a controversial phenomenon. *Nat Rev Neurosci* 2012;13:859–66.
- [165] Morena M, Patel S, Bains JS, Hill MN. Neurobiological interactions between stress and the endocannabinoid system. *Neuropsychopharmacology* 2016;41:80–102.
- [166] Mulpuri Y, Marty VN, Munier JJ, Mackie K, Schmidt BL, Seltzman HH, Spigelman I. Synthetic peripherally-restricted cannabinoid suppresses

- chemotherapy-induced peripheral neuropathy pain symptoms by CB1 receptor activation. *Neuropharmacology* 2018;139:85–97.
- [167] Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for cannabinoids. *Nature* 1993;365:61–5.
- [168] Myers AM, Siegle PB, Foss JD, Tuma RF, Ward SJ. Single and combined effects of plant-derived and synthetic cannabinoids on cognition and cannabinoid-associated withdrawal signs in mice. *Br J Pharmacol* 2019;176:1552–67.
- [169] Nackley AG, Suplita RL II, Hohmann AG. A peripheral cannabinoid mechanism suppresses spinal fos protein expression and pain behavior in a rat model of inflammation. *Neuroscience* 2003;117:659–70.
- [170] Nackley AG, Zvonok AM, Makriyannis A, Hohmann AG. Activation of cannabinoid CB2 receptors suppresses C-fiber responses and windup in spinal wide dynamic range neurons in the absence and presence of inflammation. *J Neurophysiol* 2004;92:3562–74.
- [171] Naef M, Curatolo M, Petersen-Felix S, Arendt-Nielsen L, Zbinden A, Brenneisen R. The analgesic effect of oral delta-9-tetrahydrocannabinol (THC), morphine, and a THC-morphine combination in healthy subjects under experimental pain conditions. *PAIN* 2003;105:79–88.
- [172] Naef M, Russmann S, Petersen-Felix S, Brenneisen R. Development and pharmacokinetic characterization of pulmonary and intravenous delta-9-tetrahydrocannabinol (THC) in humans. *J Pharm Sci* 2004;93:1176–84.
- [173] Nahler G, Grotenhermen F, Zuardi AW, Crippa JAS. A conversion of oral cannabidiol to delta9-tetrahydrocannabinol seems not to occur in humans. *Cannabis Cannabinoid Res* 2017;2:81–6.
- [174] Naidu PS, Booker L, Cravatt BF, Lichtman AH. Synergy between enzyme inhibitors of fatty acid amide hydrolase and cyclooxygenase in visceral nociception. *J Pharmacol Exp Ther* 2009;329:48–56.
- [175] Navarrete F, Rodriguez-Arias M, Martin-Garcia E, Navarro D, Garcia-Gutierrez MS, Aguilar MA, Aracil-Fernandez A, Berbel P, Minarro J, Maldonado R, Manzanares J. Role of CB2 cannabinoid receptors in the rewarding, reinforcing, and physical effects of nicotine. *Neuropsychopharmacology* 2013;38:2515–24.
- [176] Neelakantan H, Tallarida RJ, Reichenbach ZW, Tuma RF, Ward SJ, Walker EA. Distinct interactions of cannabidiol and morphine in three nociceptive behavioral models in mice. *Behav Pharmacol* 2015;26:304–14.
- [177] Negus SS, Bilsky EJ, Do Carmo GP, Stevenson GW. Rationale and methods for assessment of pain-depressed behavior in preclinical assays of pain and analgesia. *Methods Mol Biol* 2010;617:79–91.
- [178] Nyilas R, Gregg LC, Mackie K, Watanabe M, Zimmer A, Hohmann AG, Katona I. Molecular architecture of endocannabinoid signaling at nociceptive synapses mediating analgesia. *Eur J Neurosci* 2009;29:1964–78.
- [179] O'Sullivan SE. Cannabinoids go nuclear: evidence for activation of peroxisome proliferator-activated receptors. *Br J Pharmacol* 2007;152:576–82.
- [180] Ohlsson A, Lindgren JE, Wahlen A, Agurell S, Hollister LE, Gillespie HK. Plasma delta-9 tetrahydrocannabinol concentrations and clinical effects after oral and intravenous administration and smoking. *Clin Pharmacol Ther* 1980;28:409–16.
- [181] Ohno-Shosaku T, Maejima T, Kano M. Endogenous cannabinoids mediate retrograde signals from depolarized postsynaptic neurons to presynaptic terminals. *Neuron* 2001;29:729–38.
- [182] Otrubova K, Ezzili C, Boger DL. The discovery and development of inhibitors of fatty acid amide hydrolase (FAAH). *Bioorg Med Chem Lett* 2011;21:4674–85.
- [183] Pacher P, Batkai S, Kunos G. The endocannabinoid system as an emerging target of pharmacotherapy. *Pharmacol Rev* 2006;58:389–462.
- [184] Pacher P, Kunos G. Modulating the endocannabinoid system in human health and disease—successes and failures. *FEBS J* 2013;280:1918–43.
- [185] Parker LA, Mechoulam R, Schlievert C. Cannabidiol, a non-psychoactive component of cannabis and its synthetic dimethylheptyl homolog suppress nausea in an experimental model with rats. *Neuroreport* 2002;13:567–70.
- [186] Pascual D, Goicoechea C, Suardiaz M, Martin MI. A cannabinoid agonist, WIN 55,212-2, reduces neuropathic nociception induced by paclitaxel in rats. *PAIN* 2005;118:23–34.
- [187] Patel S, Hill MN, Cheer JF, Wotjak CT, Holmes A. The endocannabinoid system as a target for novel anxiolytic drugs. *Neurosci Biobehav Rev* 2017;76:56–66.
- [188] Paudel KS, Hammell DC, Agu RU, Valiveti S, Stinchcomb AL. Cannabidiol bioavailability after nasal and transdermal application: effect of permeation enhancers. *Drug Dev Ind Pharm* 2010;36:1088–97.
- [189] Paunescu H, Coman OA, Coman L, Ghita I, Georgescu SR, Draghia F, Fulga I. Cannabinoid system and cyclooxygenases inhibitors. *J Med Life* 2011;4:11–20.
- [190] Percie du Sert N, Rice AS. Improving the translation of analgesic drugs to the clinic: animal models of neuropathic pain. *Br J Pharmacol* 2014;171:2951–63.
- [191] Pertwee RG. Pharmacology of cannabinoid CB1 and CB2 receptors. *Pharmacol Ther* 1997;74:129–80.
- [192] Pertwee RG. Cannabinoid receptors and pain. *Prog Neurobiol* 2001;63:569–611.
- [193] Pertwee RG. *Hanbook of Cannabis*. Oxford, United Kingdom: Oxford University Press, 2016.
- [194] Petitot F, Jeantaud B, Bertrand P, Imperato A. Cannabinoid penetration into mouse brain as determined by ex vivo binding. *Eur J Pharmacol* 1999;374:417–21.
- [195] Philpott HT, O'Brien M, McDougall JJ. Attenuation of early phase inflammation by cannabidiol prevents pain and nerve damage in rat osteoarthritis. *PAIN* 2017;158:2442–51.
- [196] Piomelli D, Hohmann AG, Seybold V, Hammock BD. A lipid gate for the peripheral control of pain. *J Neurosci* 2014;34:15184–91.
- [197] Piomelli D, Tarzia G, Duranti A, Tontini A, Mor M, Compton TR, Dasse O, Monaghan EP, Parrott JA, Putman D. Pharmacological profile of the selective FAAH inhibitor KDS-4103 (URB597). *CNS Drug Rev* 2006;12:21–38.
- [198] Pryor GT, Husain S, Mitoma C. Influence of fasting on the absorption and effects of delta9-tetrahydrocannabinol after oral administration in sesame oil. *Pharmacol Biochem Behav* 1977;6:331–41.
- [199] Rahn EJ, Deng L, Thakur GA, Vemuri K, Zvonok AM, Lai YY, Makriyannis A, Hohmann AG. Prophylactic cannabinoid administration blocks the development of paclitaxel-induced neuropathic nociception during analgesic treatment and following cessation of drug delivery. *Mol Pain* 2014;10:27.
- [200] Rani Sagar D, Burston JJ, Woodhams SG, Chapman V. Dynamic changes to the endocannabinoid system in models of chronic pain. *Philos Trans R Soc Lond B Biol Sci* 2012;367:3300–11.
- [201] Rice AS, Cimino-Brown D, Eisenach JC, Kontinen VK, Lacroix-Fralish ML, Machin I, Preclinical Pain C, Mogil JS, Stohr T. Animal models and the prediction of efficacy in clinical trials of analgesic drugs: a critical appraisal and call for uniform reporting standards. *PAIN* 2008;139:243–7.
- [202] Rice ASC, Finnerup NB, Kemp HI, Currie GL, Baron R. Sensory profiling in animal models of neuropathic pain: a call for back-translation. *PAIN* 2018;159:819–24.
- [203] Rice ASC, Morland R, Huang W, Currie GL, Sena ES, Macleod MR. Transparency in the reporting of in vivo pre-clinical pain research: the relevance and implications of the ARRIVE (Animal Research: reporting in Vivo Experiments) guidelines. *Scand J Pain* 2013;4:58–62.
- [204] Richardson JD, Kilo S, Hargreaves KM. Cannabinoids reduce hyperalgesia and inflammation via interaction with peripheral CB1 receptors. *PAIN* 1998;75:111–19.
- [205] Roche M, Finn DP. Brain CB(2) receptors: implications for neuropsychiatric disorders. *Pharmaceuticals (Basel)*. 2010;3:2517–53.
- [206] Sagar DR, Gaw AG, Okine BN, Woodhams SG, Wong A, Kendall DA, Chapman V. Dynamic regulation of the endocannabinoid system: implications for analgesia. *Mol Pain* 2009;5:59.
- [207] Sagar DR, Jhaveri MD, Richardson D, Gray RA, de Lago E, Fernandez-Ruiz J, Barrett DA, Kendall DA, Chapman V. Endocannabinoid regulation of spinal nociceptive processing in a model of neuropathic pain. *Eur J Neurosci* 2010;31:1414–22.
- [208] Sain NM, Liang A, Kane SA, Urban MO. Antinociceptive effects of the non-selective cannabinoid receptor agonist CP 55,940 are absent in CB1(-/-) and not CB2(-/-) mice in models of acute and persistent pain. *Neuropharmacology* 2009;57:235–41.
- [209] Schuster RM, Potter K, Vandrey R, Hareli M, Gilman J, Schoenfeld D, Ewins AE. Urinary 11-nor-9-carboxy-tetrahydrocannabinol elimination in adolescent and young adult cannabis users during one month of sustained and biochemically-verified abstinence. *J Psychopharmacol* 2020;34:197–210.
- [210] Sholler DJ, Strickland JC, Spindle TR, Weerts EM, Vandrey R. Sex differences in the acute effects of oral and vaporized cannabis among healthy adults. *Addict Biol*:e129682020. doi: 10.1111/adb.12966.
- [211] Slivicki RA, Iyer V, Mali SS, Garai S, Thakur GA, Crystal JD, Hohmann AG. Positive allosteric modulation of CB1 cannabinoid receptor signaling enhances morphine antinociception and attenuates morphine tolerance without enhancing morphine-induced dependence or reward. *Front Mol Neurosci* 2020;13:54.
- [212] Slivicki RA, Saberi SA, Iyer V, Vemuri VK, Makriyannis A, Hohmann AG. Brain-permeant and -impermeant inhibitors of fatty acid amide hydrolase synergize with the opioid analgesic morphine to suppress chemotherapy-induced neuropathic nociception without enhancing effects of morphine on gastrointestinal transit. *J Pharmacol Exp Ther* 2018;367:551–63.

- [213] Slivicki RA, Xu Z, Mali SS, Hohmann AG. Brain permeant and impermeant inhibitors of fatty-acid amide hydrolase suppress the development and maintenance of paclitaxel-induced neuropathic pain without producing tolerance or physical dependence in vivo and synergize with paclitaxel to reduce tumor cell line viability in vitro. *Pharmacol Res* 2019;142:267–82.
- [214] Smith PB, Compton DR, Welch SP, Razdan RK, Mechoulam R, Martin BR. The pharmacological activity of anandamide, a putative endogenous cannabinoid, in mice. *J Pharmacol Exp Ther* 1994;270:219–27.
- [215] Soliman N, Vollert J, Sena C, Liao J, Madeod M, Thomas J, Hohmann AG, Haroutounian S, Krane E, Alaverdyan H, Barakat A, Barthlow T, Harris Bozer AL, Davidson A, Diaz-delCastillo M, Dolgorukova A, Ferdousi M, Healy C, Hong N, Hopkins M, James A, Leake HB, Malewicz NM, Mansfield M, Mardon AK, Mattimoe D, McLoone DP, Noes-Holt G, Pogatzki-Zahn EM, Power E, Pradier B, Romanos-Sirakis E, Segelcke A, Segelcke D, Vinagre R, Yanes JA, Zhang J, Zhang XY, Finn DP, Rice ASC. Systematic Review and Meta-analysis of cannabis-based medicines, cannabinoids and endocannabinoid system modulators tested for antinociceptive effects in animal models of injury-related or pathological persistent pain. *PAIN* 2021;162(S1):S26–S44.
- [216] Spencer S, Neuhofer D, Chioma VC, Garcia-Keller C, Schwartz DJ, Allen N, Scofield MD, Ortiz-Ithier T, Kalivas PW. A model of delta(9)-tetrahydrocannabinol self-administration and reinstatement that alters synaptic plasticity in nucleus accumbens. *Biol Psychiatry* 2018;84:601–10.
- [217] Starowicz K, Finn DP. Cannabinoids and pain: sites and mechanisms of action. *Adv Pharmacol* 2017;80:437–75.
- [218] Stella N, Schweitzer P, Piomelli D. A second endogenous cannabinoid that modulates long-term potentiation. *Nature* 1997;388:773–8.
- [219] Sugiura T, Kishimoto S, Oka S, Gokoh M. Biochemistry, pharmacology and physiology of 2-arachidonoylglycerol, an endogenous cannabinoid receptor ligand. *Prog Lipid Res* 2006;45:405–46.
- [220] Sugiura T, Kondo S, Sukagawa A, Nakane S, Shinoda A, Itoh K, Yamashita A, Waku K. 2-Arachidonoylglycerol: a possible endogenous cannabinoid receptor ligand in brain. *Biochem Biophys Res Commun* 1995;215:89–97.
- [221] Taylor L, Crockett J, Tayo B, Morrison G. A phase 1, open-label, parallel-group, single-dose trial of the pharmacokinetics and safety of cannabidiol (CBD) in subjects with mild to severe hepatic impairment. *J Clin Pharmacol* 2019;59:1110–19.
- [222] Tham M, Yilmaz O, Alaverdashvili M, Kelly MEM, Denovan-Wright EM, Laprairie RB. Allosteric and orthosteric pharmacology of cannabidiol and cannabidiol-dimethylheptyl at the type 1 and type 2 cannabinoid receptors. *Br J Pharmacol* 2019;176:1455–69.
- [223] Thors L, Burston JJ, Alter BJ, McKinney MK, Cravatt BF, Ross RA, Pertwee RG, Gereau RW IV, Wiley JL, Fowler CJ. Biochanin A, a naturally occurring inhibitor of fatty acid amide hydrolase. *Br J Pharmacol* 2010;160:549–60.
- [224] Tjolsen A, Berge OG, Hunskaar S, Rosland JH, Hole K. The formalin test: an evaluation of the method. *PAIN* 1992;51:5–17.
- [225] Tseng AH, Harding JW, Craft RM. Pharmacokinetic factors in sex differences in Delta 9-tetrahydrocannabinol-induced behavioral effects in rats. *Behav Brain Res* 2004;154:77–83.
- [226] Tsou K, Lowitz KA, Hohmann AG, Martin WJ, Hathaway CB, Bereiter DA, Walker JM. Suppression of noxious stimulus-evoked expression of Fos protein-like immunoreactivity in rat spinal cord by a selective cannabinoid agonist. *Neuroscience* 1996;70:791–8.
- [227] Ueda N, Tsuboi K, Uyama T, Ohnishi T. Biosynthesis and degradation of the endocannabinoid 2-arachidonoylglycerol. *Biofactors* 2011;37:1–7.
- [228] Uhelski ML, Khasabova IA, Simone DA. Inhibition of anandamide hydrolysis attenuates nociceptor sensitization in a murine model of chemotherapy-induced peripheral neuropathy. *J Neurophysiol* 2015;113:1501–10.
- [229] Valveti S, Hammell DC, Earles DC, Stinchcomb AL. Transdermal delivery of the synthetic cannabinoid WIN 55,212-2: in vitro/in vivo correlation. *Pharm Res* 2004;21:1137–45.
- [230] van der Worp HB, Howells DW, Sena ES, Porritt MJ, Rewell S, O'Collins V, Madeod MR. Can animal models of disease reliably inform human studies? *PLoS Med* 2010;7:e1000245.
- [231] Voelkl B, Vogt L, Sena ES, Wurbel H. Reproducibility of preclinical animal research improves with heterogeneity of study samples. *PLoS Biol* 2018;16:e2003693.
- [232] Wagenlehner FME, van Till JWO, Houbiers JGA, Martina RV, Cerneus DP, Melis J, Majek A, Vjaters E, Urban M, Ramonas H, Shoskes DA, Nickel JC. Fatty acid amide hydrolase inhibitor treatment in men with chronic prostatitis/chronic pelvic pain syndrome: an adaptive double-blind, randomized controlled trial. *Urology* 2017;103:191–7.
- [233] Wang J. Glial endocannabinoid system in pain modulation. *Int J Neurosci* 2019;129:94–100.
- [234] Welch SP, Huffman JW, Lowe J. Differential blockade of the antinociceptive effects of centrally administered cannabinoids by SR141716A. *J Pharmacol Exp Ther* 1998;287:1301–8.
- [235] Whiteside GT, Adedoyin A, Leventhal L. Predictive validity of animal pain models? A comparison of the pharmacokinetic-pharmacodynamic relationship for pain drugs in rats and humans. *Neuropharmacology* 2008;54:767–75.
- [236] Wiese BM, Liktov-Busa E, Levine A, Couture SA, Nikas SP, Ji L, Liu Y, Mackie K, Makriyannis A, Largent-Milnes TM, Vanderah TW. Cannabinoid-2 agonism with AM2301 mitigates morphine-induced respiratory depression. *Cannabis Cannabinoid Res* 2020. doi:10.1089/can.2020.0076. Epub ahead of print.
- [237] Wilkerson JL, Ghosh S, Mustafa M, Abdullah RA, Niphakis MJ, Cabrera R, Maldonado RL, Cravatt BF, Lichtman AH. The endocannabinoid hydrolysis inhibitor SA-57: intrinsic antinociceptive effects, augmented morphine-induced antinociception, and attenuated heroin seeking behavior in mice. *Neuropharmacology* 2017;114:156–67.
- [238] Wilkerson JL, Milligan ED. The central role of glia in pathological pain and the potential of targeting the cannabinoid 2 receptor for pain relief. *ISRN Anesthesiol* 2011;2011.
- [239] Wilson RI, Nicoll RA. Endogenous cannabinoids mediate retrograde signalling at hippocampal synapses. *Nature* 2001;410:588–92.
- [240] Woodhams SG, Chapman V, Finn DP, Hohmann AG, Neugebauer V. The cannabinoid system and pain. *Neuropharmacology* 2017;124:105–20.
- [241] Woodhams SG, Sagar DR, Burston JJ, Chapman V. The role of the endocannabinoid system in pain. *Handbook Exp Pharmacol* 2015;227:119–43.
- [242] Wu J, Hocevar M, Bie B, Foss JF, Naguib M. Cannabinoid type 2 receptor system modulates paclitaxel-induced microglial dysregulation and central sensitization in rats. *J Pain* 2019;20:501–14.
- [243] Xiong W, Cui T, Cheng K, Yang F, Chen SR, Willenbring D, Guan Y, Pan HL, Ren K, Xu Y, Zhang L. Cannabinoids suppress inflammatory and neuropathic pain by targeting alpha3 glycine receptors. *J Exp Med* 2012;209:1121–34.
- [244] Xu C, Chang T, Du Y, Yu C, Tan X, Li X. Pharmacokinetics of oral and intravenous cannabidiol and its antidepressant-like effects in chronic mild stress mouse model. *Environ Toxicol Pharmacol* 2019;70:103202.
- [245] Yuill MB, Hale DE, Guindon J, Morgan DJ. Anti-nociceptive interactions between opioids and a cannabinoid receptor 2 agonist in inflammatory pain. *Mol Pain* 2017;13:1744806917728227.
- [246] Zavala CA, Thomaz AC, Iyer V, Mackie K, Hohmann AG. Cannabinoid CB2 receptor activation attenuates fentanyl-induced respiratory depression. *Cannabis Cannabinoid Res* 2020. doi: 10.1089/can.2020.0059 [Epub ahead of print].
- [247] Zgair A, Lee JB, Wong JCM, Taha DA, Aram J, Di Virgilio D, McArthur JW, Cheng YK, Hennig IM, Barrett DA, Fischer PM, Constantinescu CS, Gershkovich P. Oral administration of cannabis with lipids leads to high levels of cannabinoids in the intestinal lymphatic system and prominent immunomodulation. *Sci Rep* 2017;7:14542.
- [248] Zgair A, Wong JC, Lee JB, Mistry J, Sivak O, Wasan KM, Hennig IM, Barrett DA, Constantinescu CS, Fischer PM, Gershkovich P. Dietary fats and pharmaceutical lipid excipients increase systemic exposure to orally administered cannabis and cannabis-based medicines. *Am J Transl Res* 2016;8:3448–59.
- [249] Zhang HY, Bi GH, Li X, Li J, Qu H, Zhang SJ, Li CY, Onaivi ES, Gardner EL, Xi ZX, Liu QR. Species differences in cannabinoid receptor 2 and receptor responses to cocaine self-administration in mice and rats. *Neuropsychopharmacology* 2015;40:1037–51.
- [250] Zhu CZ, Mikusa JP, Fan Y, Hollingsworth PR, Pai M, Chandran P, Daza AV, Yao BB, Dart MJ, Meyer MD, Decker MW, Hsieh GC, Honore P. Peripheral and central sites of action for the non-selective cannabinoid agonist WIN 55,212-2 in a rat model of post-operative pain. *Br J Pharmacol* 2009;157:645–55.